A REVIEW ON FORMULATION AND EVALUATION OF MICROSPHERES

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ABSTRACT:
Multiparticulate drug delivery systems called microspheres are designed to deliver drugs to specific sites at a predetermined rate, enhance bioavailability, and achieve controlled or sustained drug administration. Polymeric waxy materials or other protective materials like natural, semi-synthetic, and synthetic polymers are used to make them. Microspheres are made of proteins or synthetic polymers and are characterised by their free-flowing, 1-1000 μm particle size. The variety of methods available for creating microspheres offers a multitude of choices for managing drug delivery and optimising the medicinal effectiveness of a particular medication. When compared to traditional dose forms, these administration methods have many benefits, such as increased patient compliance, decreased toxicity, and enhanced efficacy. These kinds of systems frequently employ macromolecules as medication carriers. In the current review, several kinds of microspheres, preparation techniques, applications, and efficiency evaluation parameters are highlighted.

KEYWORDS: Microsphere, Types of microsphere, Methods of microsphere, Application of Microspheres.

Introduction

Microspheres are defined as roughly spherical, solid particles with a diameter ranging from 1 to 1000 μm. They can also take the form of microcrystalline particles or pharmaceuticals disseminated in a particular solution. Microspheres and microcapsules are frequently used interchangeably.
Micromatrices, in which the entrapped material is diffusing throughout the microsphere matrix, and microcapsules, in which the entrapped substance is clearly encircled by distinct capsule walls, are two types of microcapsules. A drug that has been dissolved or disseminated across a particle matrix can potentially release the drug under regulated conditions when it is incorporated into solid biodegradable microspheres. They are composed of biodegradable synthetic polymers and modified natural products, as well as polymeric, waxy, and other protective components.

Medication that enters the bloodstream through the gastrointestinal tract (GIT) and has a brief half-life is promptly eliminated by the blood circulation. In order to prevent this issue, oral sustained release (OSR) or controlled release (CR) has also been developed. OSR or CR will gradually release the drug from the GIT and maintain a consistent dose in the plasma for an extended amount of time. A dose formulation that achieves the necessary plasma therapeutic drug concentration and stays stable over the course of the treatment is considered appropriate. This can be accomplished by administering a conventional dosage form at a predetermined frequency and dose.

An advantage They're not Microcarriers larger than nanoparticles move into the interstitium, where they act locally, over the 100 nm range that the lymph carries. Most likely, hazardous substances can be conveyed. Encapsulated, the dried microparticles may be referred to as solids instead of liquid. Since the intake dose is given in a number of small, distinct multiparticulate particles that each hold and release a portion of the dosage, the failure of one subunit does not impact the dosage failure as a whole.

When applied topically, microparticles that facilitate the drug's release into the skin work to keep the medication localised at the application site and prevent it from entering the bloodstream needlessly.

They serve as a reservoir that gradually releases an active component to maintain an appropriate concentration of medication items in the skin while reducing undesirable side effects. As a result, there are fewer cycles of excessive and insufficient medication.
In the treatment of infectious diseases, it is particularly pertinent to the decrease of antibiotic resistance. Additionally, these distribution channels can improve product safety or integration into the right vehicles.

**Advantages:**

1. A smaller microsphere enhances the surface area and, as a result, the potency of the poorly soluble substance.

2. It is possible to lower dosage frequency and side effects.

3. A rise in patient adherence

4. Drugs enclosed in polymers shield them from enzymatic cleavage, protecting them from a range of enzymes.

5. Promotes better bioavailability.

6. Reduced gastric irritation is possible.

7. The biological half-life is improvable.

8. It is possible to lower first pass metabolism.

9. The drug's unpleasant taste and aroma can be covered up.

10. have a long-lasting and consistent therapeutic impact.

11. Their smaller size and spherical form allow them to be injected into the body.

12. Convert a liquid into a solid and eliminate the bad flavour.

13. Degradable microspheres with controlled release distribution are being employed to control drug price releases while lowering toxicity and the pain associated with repeated injections.

**Disadvantages:**

1. One of the drawbacks is the altered releases from the formulations.

2. The rate at which the regulated dose is released, which varies depending on a lot of factors like food and amounts transferred via the gastrointestinal tract.

3. Discharge rates varying from one dosage to the next.

4. Because controlled release formulations usually have a larger dose load, any deficiency in the drug substance's release qualities could lead to E. Potentially harmful.

5. Chewing or breaking these dosage forms is prohibited

6. There is less reproducibility.

7. In contrast to traditional preparations, the cost of ingredients and processing is substantial.
8. The stability of core particles may be impacted by changes in process factors such as temperature, pH, solvent addition, and agitation/evaporation.

9. The destiny of additives and polymer matrix

**Materials utilised in the formulation of the microspheres:**

Polymers are the primary ingredients used in the formulation of the microspheres, and they are categorised as follows.

- Natural polymers
- Synthetic polymers

A. There are two categories of synthetic polymers.

a) Polymethyl methacrylate (PMMA), acryllein glycidyl methacrylate, and epoxy polymers are examples of non-biodegradable polymers.

b) Biodegradable polymers: Anhydrides, poly alkyl cyano acrylates, lactides, and glycolides and their copolymers

B. Natural polymers: They come from a variety of sources, including proteins, carbs, and carbohydrates that have undergone chemical modification. Additionally, proteins like collagen, gelatin, and albumin are utilised by them. Carbohydrates such as poly dextran and poly starch, as well as carbohydrates that have been chemically altered, such as agarose, carrageenan, chitosan, and starch

**Microsphere types:**

1) **Bio-adhesive microspheres:** Adhesion is defined as the ability to stick to a membrane through the usage of the water-soluble polymer's sticking characteristics. The bioadhesion property of some polymers, which stick to one another when hydrated, is used by the bioadhesive drug delivery system to deliver medication to a particular part of the body over an extended period of time. As a result, the drug's absorption and, consequently, bioavailability are enhanced by the lowered dosage frequency, which increases patient compliance.

2) **Magnetic Microspheres:** This type of drug delivery technology is crucial since it helps to target the exact location of the illness. A smaller quantity of a medicine with magnetic targeting can take the place of the greater amount of the drug that is freely circulating. Chitosan, dextran, and other materials that are integrated into magnetic microspheres cause magnetic carriers to respond magnetically to a magnetic field. The various varieties are

Chemotherapeutic agents are delivered to liver tumours using therapeutic magnetic microspheres. This technique can also target drugs such as proteins and peptides.
3) **Diagnostic microspheres:** By producing nano-sized particles called supramagnetic iron oxides, they can be utilised to identify bowel loops from other abdominal structures and to image liver metastases.

![Fig: 2 diagnostic microsphere](image)

4) **Floating microspheres:** These microspheres stay buoyant in the stomach without influencing the rate of gastric emptying because their bulk density is lower than that of gastric fluid. If the system is floating on stomach content, the drug is released gradually at the intended rate, increasing gastric residence and fluctuating plasma concentration. Additionally, it lessens the likelihood of dosage dumping and striking. It also results in a longer-lasting therapeutic impact, which lowers the frequency of dose. Ketoprofen is administered using this method.

![Fig: 3 floating microsphere](image)

5) **Radioactive microspheres:** 10–30 nm-sized radio embolisation therapy microspheres are larger than capillary diameters and are tapped into the first capillary bed upon encounter. All of these conditions result in radioactive microspheres, which give significant radiation doses to the targeted locations without harming the normal surrounding tissues, because they are injected into the arteries that lead them to the tumour of interest. Unlike drug delivery systems, it operates from a radioisotope-typical
distance rather than releasing radioactivity from microspheres. The three types of radioactive microspheres are α, β, and γ emitters.

fig: 4 radioactivr microsphere

6) **Polymeric microspheres:**

There are two types of polymeric microspheres: synthetic and biodegradable.

**i. Microspheres of biodegradable polymeric material**

Since natural polymers like starch are biodegradable, biocompatible, and naturally bioadhesive, they are utilised. Because biodegradable polymers have a high degree of swelling property with aqueous medium, which results in gel formation, they have a longer residence duration when in contact with mucosal membranes. The polymer concentration and the sustained release pattern regulate the drug's release's rate and extent. This kind of microsphere has an extended residence period at the application location.

**ii. Microspheres made of synthetic polymers**

The main drawback of synthetic polymeric microspheres, despite their widespread use in clinical applications, is that they have a tendency to migrate away from the injection site, which increases the risk of embolism and subsequent organ damage. These microspheres can also be used as bulking agents, fillers, embolic particles, drug delivery vehicles, etc.

**Techniques for preparing microspheres:**

Below are several techniques for creating microspheres:

1. The method of evaporating emulsion solution
2. The cross-linking emulsion technique
3. Method of coacervation
4. The spray-dry method
5. Diffusion of emulsion and solvent
6. The multiple emulsion approach
7. Gelation of ions
8. Hydroxyl appetite microspheres (HAP) in the shape of spheres
1. The method of evaporating emulsion solution

This method involves dissolving the medication in a polymer that has already been dissolved in chloroform, and then adding the resultant solution to the aqueous phase that contains 0.2% sodium PVP as an emulsifying agent. After 500 rpm of agitation, the medicine and polymer (eudragit) were separated into fine droplets, which were subsequently collected by filtering, cleaned with demineralized water, and dried at room temperature for a whole day. The solidified microspheres were created by solvent evaporation. Using this method, aceclofenac microspheres were created.

2. Emulsion cross-linking method:

In this procedure, the medication was dissolved in an aqueous gelation solution that had been heated to 40 degrees Celsius for one hour beforehand. Drop by drop, the solution was added to the liquid paraffin while the combination was stirred at 1500 rpm for 10 minutes at 35 degrees Celsius. This produced an emulsion, which was then further stirred for 10 minutes at 15 degrees Celsius. The resulting microspheres were thus treated with 100mL of 10mm glycine solution containing 0.1%w/v of tween 80 at 37oC for 10 min to block unreacted glutaraldehyde, after which they were air dried and dispersed in 5mL of aqueous glutaraldehyde saturated toluene solution for three hours to facilitate cross-linking. Examples of this method include Gelatin A microspheres.

3. Method of coacervation

Escalation of temperature change: Ethyl cellulose was weighed and then heated to 80°C in cyclohexane while being vigorously stirred. Subsequently, the medication was finely ground and added to the aforesaid solution while being vigorously stirred. Phase separation was achieved by lowering the temperature and employing an ice bath. The product above was then air dried, cleaned twice with cyclohexane, and sieved (sieve no. 40) to produce individual microcapsules.

Coacervation without solvent addition:

This was created by dispersing the medication in a closed beaker with magnetic stirring for six hours at 500 rpm after weighing out a quantity of ethyl cellulose and dissolving it in toluene containing propylisobutylene. The stirring was then maintained for fifteen minutes. Petroleum benzoin is then used to complete phase separation.14 times while constantly stirring. The microcapsules were then cleaned with n-hexane, allowed to air dry for two hours, then baked for four hours at 50 degrees Celsius.

4. The method of spray drying

This method was employed to create polymeric-blend microspheres that were filled with the medication ketoprofen. It entails mixing the liquid coating material with the core material, spraying the combination outside to solidify the coating and quickly evaporate the solvent. To create drug-loaded microspheres, an organic solution containing polye (epsilon caprolactone) (PCL), cellulose acetate butyrate (CAB), and ketoprofen in various weight ratios was produced and sprayed under various experimental conditions. This is quick, however because of the quick drying process, it could lose crystalinity.

5. Diffusion of emulsion and solvent:

Diffusion method of emulsion and solvent Diffusion solvent diffusion technology was used to manufacture ketoprofen floating microparticles in order to increase the residence period in the colon. The drug polymer
combination was first dissolved in a 1:1 mixture of ethanol and dichloromethane. The mixture was then gradually added to a solution of sodium lauryl sulphate (SLS). For one hour, the solution was agitated at room temperature at 150 rpm using a propeller-style agitator. As a result, the generated floating microspheres were cleaned and allowed to dry at room temperature in a desiccator. The subsequent microparticles were gathered and sieved.

6. The multiple emulsion technique
This method was used to prepare indomethacin for oral controlled release medication administration. Drug powder was first dispersed in methyl cellulose solution and then emulsified in ethyl cellulose solution in ethyl acetate. After that, an aqueous medium was used to reemulsify the original emulsion. Discrete microspheres generated during this phase under optimal conditions.

7. Gelation of ions
This method was used to generate an alginate/chitosan particulate system for the release of diclofenac sodium. 1.2% (w/v) sodium alginate aqueous solution was mixed with 25% (w/v) diclofenac sodium. Stirring is continued until the entire solution is obtained, and then it is added dropwise to a solution containing acetic acid, Ca2+/Al3+, and chitosan solution. After internal gellification for 24 hours in the original solution, the produced microspheres were filtered to facilitate separation. The medication did not release in an acidic pH; nevertheless, the full release was obtained in pH 6.4–7.2.1.

8. Microspheres of hydroxyapatite (HAP) in sphere shape
This was employed in the creation of microspheres with unusual sphere morphology. O/w emulsion was used to create the microspheres, and then the solvent was evaporated. Initially, an o/w emulsion was created by dispersing the organic phase (Diclofenac sodium with 5% w/w EVA and the required quantity of HAP) into the surfactant's aqueous phase. Small droplets of the organic phase were distributed, and they remained separate droplets because the droplets were encircled by surfactant molecules, which kept them from co-solving. The DCM gradually evaporated while being stirred, and each droplet solidified to form a microsphere.

Microsphere evaluation:

1) Particle size analyzer:
In order to prevent microsphere aggregation, 50 mg of microspheres are suspended in 5 mL of distilled water with 2% w/v of tween 80. The aforementioned suspension is then sonicated in a water bath, and the particle size is expressed as volume mean diameter in micrometers.

2) Optical microscopy:
This technique uses an optical microscope (Meizer OPTIK) to measure the size of the particles. I measured under 450x (10x eye piece and 45x objective), and the results show that there are 100 particles.

3) SEM, or scanning electron microscopy:
The SEM method determines surface morphology. Using double-sided sticky tape, the microcapsules are directly placed on the SEM sample slab, covered with gold film at low pressure, and examined.

4) Swelling index:
Microspheres made of sodium alginate are characterised using this method. A wire basket is filled with several solutions (100 mL), such as distilled water, buffer solutions of pH (1.2, 4.5, and 7.4), and alginate
Aqueous microspheres (100 mg), which are stored on top of the aforesaid solutions and allowed to swell at 37 °C. As a result, weight is taken on a regular basis and soaked in filter paper to determine variations in weight fluctuation between the original weight of the microspheres and weight owing to swelling.

5) **Entrapment efficiency:**
Drug-containing microspheres (5 mg) are crushed, dissolved in distilled water for three hours with the aid of an ultrasonic stirrer, filtered, and then subjected to a UV-vis spectroscopic analysis. The ratio of real drug content to theoretical drug content is known as entrapment efficiency.

X-ray diffraction: This method can be used to assess a drug's shift in crystallinity. XRD instruments are used to examine microparticles and their constituent parts [32]. angle of scanning between 80°C and 70°C.

6) **Thermal analysis:**
Differential scanning calorimetry (DSC) can be used to analyse the microcapsule's and its components' temperatures. TGA, or thermogravimetric analysis

7) **Thermometric differential analysis (DTA):**
The sample is precisely weighed and cooked on an alumina pan at a steady temperature of 10 °C per minute while under a 40 ml/min nitrogen flow.

8) **FTIR:**
FTIR can be used to detect the drug-polymer interaction as well as the drug's degradation during microencapsulation processing.

9) **Stability studies:**
To conduct a stability study, put the microspheres in a glass container with a screw lid and store them under the following circumstances:
The following parameters apply: · Ambient humidity · Room temperature (27+/−2 oC) · Oven temperature (40+/−2 oC) · Refrigerator (5 0+/−8 oC).
The study lasted for 60 days, during which time the drug content of the microsphere was examined.

10) **Zeta potential:**
After adding chitosan with varying molecular weights to the W2 phase to create the polyelectrolyte shell, the resultant particles are measured using the zeta potential method.

**Applications:**
1. Gene transfer
2. Delivery of Ophthalmic Drugs
3. Local and intratumoral medication administration
4. Oral medication administration
5. Nasal medication administration
6. Buccal administration of medication
7. Drug distribution through the digestive tract
8. Oral medication administration
9. Delivery of drugs vaginally
10. Drug administration by transdermal application
11. Colonic medication administration
12. Delivery method with many parts
13. Using microspheres to administer vaccines
14. Using microparticulate carriers for targeting
15. Microspheres targeting mediated by monoclonal antibodies

Conclusion: -

The medication delivery technique discussed in this review article—microspheres—is superior to other drug delivery systems. In the days ahead, this innovative microsphere drug delivery system will prove to be more successful in treating cancer and other diseases such as those relating to the heart, lungs, or nervous system. The formulation of the microsphere will also prove to be more potent and effective in an in-vivo delivery system. The active pharmaceutical component and other formulation-related excipients are primarily made safe by this formulation.

References:
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