



A Review On Phytochemical And Pharmacological Studies Of *Caesulia Axillaris* Roxb

¹Aeshika Sharma, ²Anshu Gangwar, ³Vimal Kumar Singh, ⁴Zafar Akbar

^{1,2}P.G. Scholar, ^{3,4}Assistant Professor

¹Department of Pharmacy,

¹M.J.P. Rohilkhand University, Bareilly, Uttar Pradesh, India- 243006

Abstract: A comparatively small but important group of plants known for their exceptional adaptations to wetland ecosystems and unique morphological characteristics makes up the genus *Caesulia*, which belongs to the *Asteraceae* family. Native to tropical Asia, including India, Bangladesh and Nepal. *Caesulia axillaris* is one of the best-known species in its genus. It is well-known for its structural adaptations and ecological importance. Furthermore, its cultural significance is highlighted by its historical usage in medicine, where different plant components have been used for generations to cure a variety of ailments. Recent scientific research has discovered a wide range of bioactive molecules in *C. axillaris*, which contribute to its pharmacological effects. The physical features, bioactive components and therapeutic benefits of *C. axillaris* are comprehensively covered in this review. It highlights the plant's capability as a valuable source of medicine and emphasizes the need for further investigation to fully comprehend its therapeutic benefits.

Index Terms - *Caesulia axillaris*, *Asteraceae*, Wetland adaptations, Bioactive compounds, Pharmacological activities.

I. INTRODUCTION

The genus *Caesulia* is a rather tiny one, belonging to the *Asteraceae* family, but it is distinguished by its specialized adaptations to wetland environments and unique morphological traits. The herbaceous growth patterns and propensity for members of this genus to flourish in aquatic or semi-aquatic habitats distinguish them¹⁻⁵. The plant's fruit, which is meant to be spread by the wind, guarantees its spread across a wide range allowing it to persist in favorable environments. By stabilizing soil and aiding in water purification processes, the presence of *C. axillaris* in wetland ecosystems supports the general health of the environment and offers food and shelter for a variety of insects, including pollinators like bees and butterflies⁶⁻⁹. Its significance is further highlighted by its use in traditional medicine and possible pharmacological effects, calling for on-going research and conservation initiatives.

The *Asteraceae*, often known as the aster, daisy, composite or sunflower family is a very diverse and numerous groups of blooming plants. There are currently accepted over 23,000 species in the group, distributed among 1,620 genera and 12 subfamilies¹⁰⁻¹⁴. Only the *Orchidaceae* has more species than the *Asteraceae*, although it is impossible to say which of the two families is genuinely larger because we don't know the precise number of species in each. A defining feature of this family is the composite inflorescence, characterized by capitula

(flower heads) encircled by a series of protective involucral bracts^{15,16}. The family's most well-known genus, Aster, is the source of the name "*Asteraceae*". It originates from the Greek term for star and is associated to its inflorescence's star shape. Concerning the word "*Compositae*," Although it is older and still valid, it clearly alludes that the family is few angiosperm families with composite blooms¹⁷⁻¹⁹. This gives it significant ecological and economic value. In many parts of the world, the *Asteraceae* family can account for up to 10% of the plant life. Although the majority of *Asteraceae* family members are herbaceous, many are also trees, vines or bushes. The family is found all over the world with the majority of members residing in barren and semiarid zones of sub-tropical and lower calm latitudes. The *Asteraceae* are a family of plants that are economically significant. Some members sell goods like lettuce, sunflower seeds, artichokes, cooking oils, herbal teas, coffee replacements and sweetening ingredients. The horticultural community loves a number of genera, such as marigolds, pot marigolds (also known as calendula), cone flowers, numerous daisies and colourful blooms²⁰⁻²². Chrysanthemums, dahlias, zinnias, heleniums and fleabane, chrysanthemums. The Grindelia, Echinacea, yarrow and a number of other members of the *Asteraceae* family are essential to herbal medicine^{23,24}.

1.1 Geographical area

Caesulia species are well suited to changing water levels and can endure submersion and they are typically found in marshes, riverbanks and other wet environments. With *Caesulia axillaris* being one of the best known, this genus includes a number of species. This species is indigenous to tropical Asia, notably India, Bangladesh and Nepal and is well-known for its unique morphology and ecological importance²⁵. It is well-suited to its wetland habitat. Due to its structural features, such as its upright and branching stems, it can flourish in flood-prone locations and its axillary blooms allow for optimal pollination even in thick flora. The only species in the monotypic genus *Caesulia* is *Caesulia axillaris*, sometimes known as the pink node flower. It may grow on damp ground or float in water bodies. It is not often weedy but in certain places, it grows in rice fields²⁶.

1.2 Agriculture importance

Groundnuts are a low-cost, nutrient-dense source of fat, protein and vitamins. The most important are India, China, the United States and West Africa, but they are cultivated commercially in the majority of the world's tropical and subtropical regions. Around 10% of all food grains in India are lost after harvest due to unscientific storage, pests, rodents, microbes and other factors²⁷⁻³⁰. The main financial harm brought on by grain-infesting insects is not always the real substance they eat; it is also the amount that their faces contaminate, rendering it unsuitable for human use. There are around 500 different insect species that have been linked to stored grain goods. Economic losses are caused by about 100 different species of insect pests of stored goods³¹⁻³⁴.

1.3 Medicinal use

In tropical and subtropical regions, herbal raw materials are especially vulnerable to fungal infection during post-harvest treatment and storage³⁵⁻³⁸. The majority of these fungi are toxic, producing micro toxin, therapy and impacting the quality of the herbal raw materials as well as the herbal composition^{39,40}. As well as antimicrobial activity. Researchers from all over the globe have conducted extensive studies with favorable outcomes on the effects of plant extracts on bacteria and fungi. The antibacterial activity of various plants was documented. Against *B. cereus*, *S. epidermis*, *S. aureus* etc⁴¹. Antioxidants serve as a storehouse of antibacterial, antifungal, anti-inflammatory and anti-carcinogenic effects. A variety of plant substances are thought to have antifungal properties without harming people or animals^{42,43}. The anti-microbial activity of *Caesulia axillaris* Roxb against pathogens in laboratory conditions has been studied by taking into account their folk medicinal properties.

In recent years, several antimicrobials derived from microorganisms have been shown to be effective against dermatophytes⁴⁴⁻⁴⁶. the plants that are empirically employed to treat skin disorders. These medications validity may be tested in several areas of fungal pathology. Regrettably, there is not much scientific and methodical study done on green plants to determine their antifungal effectiveness against ringworm^{47,48}.

Whole plant paste with camphor and mustard oil is applied on chest and throat to cure cold, cough and nasal congestion. Paste of the inflorescence with black pepper and cow milk is given to cure dysentery. Whole plant extract is given to cure malaria⁴⁹.

II. PLANT PROFILE

The genus *Caesulia* is part of the family *Asteraceae* and is a relatively small genus, yet it is notable for its special adaptations to wetland ecosystems and its unique morphological characteristics. Species within this genus are identified by their herbaceous growth forms and their ability to flourish in aquatic or semi-aquatic settings. Usually located in marshes, along riverbanks and in other saturated regions, *Caesulia* species are adapted to variable water levels and can withstand submersion. This genus comprises several species, with *Caesulia axillaris* being among the most recognized. This species is indigenous to tropical areas in Asia, especially in nations such as India, Bangladesh and Nepal is celebrated for its unique morphology and ecological importance^{50,51}. It is highly adapted to its wetland habitat. Its structural features, which include erect and branched stems, enable it to prosper in regions susceptible to flooding, while its axillary flowers promote effective pollination even amidst dense foliage⁵².



Figure 1. *Caesulia axillaris* Roxb



Figure 2. Flower of *Caesulia axillaris*

Table 1. Toxiconomical classification of *Caesulia axillaris* Roxb

Family	<i>Asteraceae</i>
Order	Asterales
Species	<i>Caesulia axillaris</i> Roxb
Kingdom	Plantae
Genus	<i>Caesulia</i> Roxb
Class	Magnoliopsida
Clade	Angiosperms

APPEARANCE^{53,54}

Common name- Pink node flower, Kala Maka, Erra gobbi, Gathila.

Habit: Glabrous, erect or decumbent herb.

Stem: 15-45 cm tall.

Leaves: 5-15 cm long, lanceolate-oblong or narrowly lanceolate, semi-amplexicaul.

Flowers: Light blue, lilac or white in globose, compound heads.

Fruits: Achenes obovoid, dark brown, ribbed, flattened.

2.1 Description

C. axillaris is a yearly herb, flourishes in wetland environments such as marshes, riverbanks and other saturated regions. It is highly suited to these surroundings and plays a crucial role in the local ecosystem⁵⁵⁻⁵⁸ (Figure 1).

Stems: The stems are upright and branched. This branching usually occurs from the base or lower portions of the stem, giving the plant a somewhat bushy look. Plants attain heights of up to 45 cm although they may differ based on specific growing conditions and environmental influences. They are smooth and frequently display a green hue, sometimes with a slight reddish tint near the base.

Leaves: The leaves are positioned alternately along the stem. They are lanceolate to elliptic, offering a streamlined and effective shape for photosynthesis. Leaf margins are entire but may occasionally be slightly serrated, contributing to their variability.

Flowers: The flowers are generally purple to blue, which can vary slightly in tone. Each flower is tubular and small, typical of many members of the *Asteraceae* family. The tubular form is made up of numerous tiny florets. Flowers are clustered in axillary formations, meaning they arise from the axils (Figure 2).

Fruits: The fruit is a tiny, single-seeded achene. An achene is a kind of simple dry fruit that remains closed at maturity and does not open to free the seed.

2.2 Traditional use

Caesulia axillaris Roxb., commonly known as 'Kala-Maka', is a herbaceous plant belonging to the *Asteraceae* family. In traditional medicine, particularly in certain regions of India, its leaves are utilized for promoting hair health. A common practice involves boiling fresh, healthy leaves in coconut oil to create a preparation that is applied to the scalp to encourage healthy and rapid hair growth⁵⁹.

Additionally, Members of the *Asteraceae* family, such as *Caesulia axillaris* are recognized for producing secondary metabolites like phenolics, flavonoids and triterpenes. These bioactive compounds are associated with several pharmacological effects, including antioxidant, anti-inflammatory and anti-microbial activities⁶⁰⁻⁶².

There has been a growth in interest in many traditional natural products in order to identify their phytochemical and anti-microbial properties. It has been proposed that aqueous and ethanol extracts from plants used in allopathic treatments are potential sources of anti-viral and anti-microbial compounds. This results in the discovery of anti-microbial potential among the native and wild flora. In traditional Indian medicine, the *Asteraceae* family's *C. axillaris* Roxb is known to treat goiter and baldness. A common weed that thrives in Indian rice fields. It has been reported that its essential oil is effective against several insect pests that cause food commodity deterioration. *Axillari* has undergone testing to see how well it works at inhibiting aflatoxin B1 and against the fungus that breaks down the raw ingredients of herbs. Additionally, animal trials have been conducted to assess the oil's safety profile in order to determine its effectiveness as a preservative for herbal raw materials. But in this study, the chemically defined oil of *C. axillaris* is analyzed for the first time. The primary goal of the current study was to identify the fungi that cause the biodegradation of the stored raw materials⁶³⁻⁶⁶.

III. PHYTOCHEMICAL STUDIES

C. axillaris is rich in various bioactive compounds that contribute to its pharmacological properties. These compounds include phenolic compounds, flavonoids, alkaloids, terpenoids, saponins and essential oils, each playing a role in the plant's antioxidant, anti-inflammatory, anti-microbial, analgesic and gastro-protective effects. The on-going study of these bioactive compounds continues to reveal the potential therapeutic benefits of *C. axillaris*. *Caesulia axillaris* Roxb contains several chemical constituents identified through phytochemical analyses⁶⁷. The notable compounds include:

3.1 γ -Asarone: A phenyl derivative constituting approximately 63.09% of the essential oil extracted from the leaves⁶⁸⁻⁷⁰.

3.2 Limonene: A monoterpenoid making up about 30.85% of the leaf essential oil⁷¹.

3.3 Flavonoids & Phenols: Present in various parts of the plant, including leaves, stems, and seeds, across different solvent extractions⁷²⁻⁷⁴.

3.4 Steroids: Detected in leaves, stems and seeds, with presence varying depending on the solvent used for extraction⁷⁵.

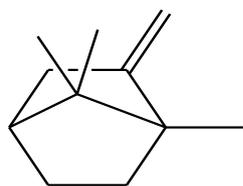
3.5 Tannins: Identified in certain plant parts, with their presence varying based on the solvent system employed during extraction⁷⁶.

Some secondary metabolite also presents such as Alkaloid, Fats & Oil, Vitamins, Amino acid, Glycoside⁷⁸⁻⁷⁹.

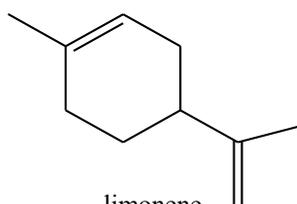
3.6 Composition of the leaf oil *Caesulia axillaris* Roxb⁶⁷⁻⁷⁹

Table 1. a= retention time, b= MS, c= 1HNMR, d=IR

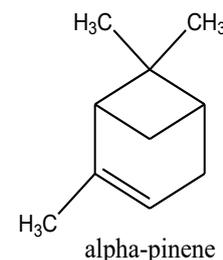
Constituents of leaf oil	Method of identification
α -pinene	a,b
camphene	a,b
limonene	a,b,c,d
p-cymene	a,b,c
terpinen-4-ol	a,b,c
p-cyrnen-7-ol	a
terpinen-4-yl acetate	a,b
α -terpineol	a,b
p-menth-8-en-1 β ,2 α -diol	b,c,d
eugenol	a,b
γ -asarone	a,b,c,d
myristicin	a,b
2,4,5-trimethoxy benzaldehyde	a,b,c,d
2-methoxy-4,5-methylenedioxy cinnamyl alcohol	b,c,d
terpinolene	a,b



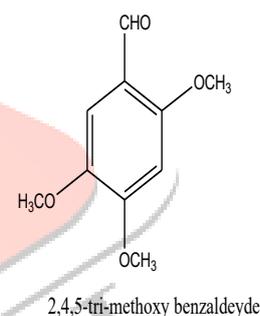
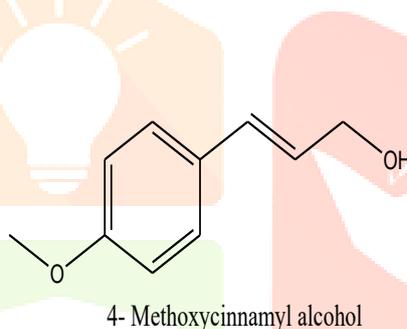
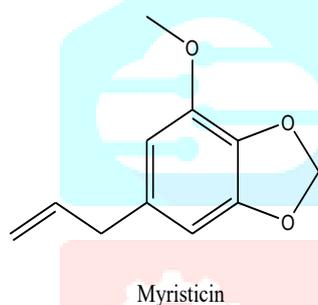
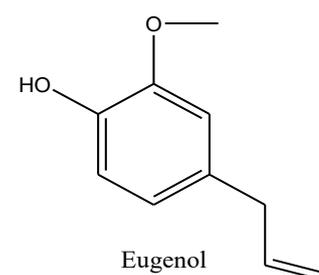
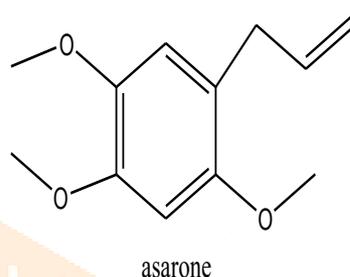
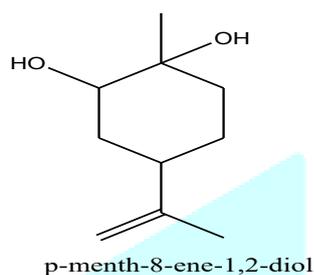
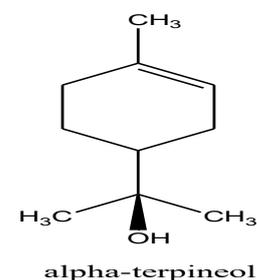
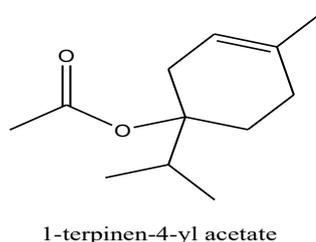
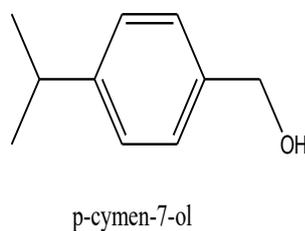
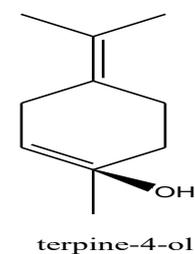
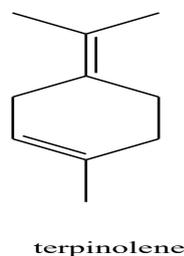
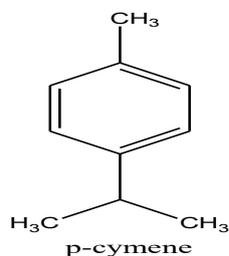
camphor



limonene



alpha-pinene



IV. PHARMACOLOGICAL STUDIES

- **Anti-microbial Activity:** Extracts from different parts of *C. axillaris* have demonstrated antimicrobial properties. Specifically, the leaves exhibited notable antifungal activity at a 2% concentration. These findings suggest that the choice of solvent and concentration plays a crucial role in the effectiveness of extraction and consequently the antimicrobial potential of the plant materials^{80,81}.
- **Antioxidant Activity:** The plant contains various phenolic compounds, flavonoids and other antioxidants. These compounds help neutralize free radicals, thereby reducing oxidative stress and potentially lowering the risk of chronic diseases such as cancer and cardiovascular diseases⁸²⁻⁸⁴.
- **Antifungal and Antiaflatoxigenic Efficacy:** The essential oil of *C. axillaris* has been evaluated for its antifungal and antiaflatoxigenic properties against fungi that deteriorate herbal raw materials. The oil exhibited significant antifungal activity and also demonstrated the ability to inhibit aflatoxin production by *Aspergillus flavus*, highlighting its potential as a natural preservative⁸⁵⁻⁸⁷.
- **Anti-inflammatory Activity:** The presence of anti-inflammatory compounds, such as flavonoids and saponins has been noted. These compounds inhibit the production of pro-inflammatory cytokines and enzymes such as cyclooxygenase (COX), which play a role in the inflammatory process^{88,89}.
- **Antipyretic Activity:** The antipyretic effect is attributed to compounds such as flavonoids and saponins. These compounds are believed to interfere with the body's production of prostaglandins, which are involved in the fever response⁹⁰⁻⁹¹.
- **Analgesic Activity:** Flavonoids and other compounds in the plant contribute to its analgesic properties. The compounds likely act by inhibiting pain pathways and reducing the perception of pain⁹²⁻⁹⁴.

- **Gastro protective Activity:** The presence of compounds with protective effects on the gastrointestinal tract has been noted. These compounds may help to strengthen the mucosal barrier, reduce gastric acid secretion or exhibit antioxidant activity to protect the lining of the stomach^{95,96}.

V. CONCLUSION

Caesulia axillaris exemplifies the remarkable synergy between ecological adaptation and medicinal utility within the genus *Caesulia*. Its structural resilience in wetland habitats, coupled with its rich pharmacological profile, positions it as a multifaceted resource worthy of conservation and exploration. Traditional knowledge of its medicinal properties finds contemporary validation through scientific inquiry, revealing various bioactive compounds with diverse therapeutic potentials. The antioxidant, anti-inflammatory, antimicrobial, analgesic, antipyretic and gastro-protective activities attributed to *C. axillaris* underscore its relevance in modern pharmacology. As research progresses, the elucidation of its bioactive constituents and pharmacological mechanisms opens avenues for the development of novel therapeutic agents.

VI. ACKNOWLEDGEMENT

We would like to express our sincere gratitude to God and Head of Département of Pharmacy M.J.P Rohilkhand University Bareilly, Uttar Pradesh, India. We are also thankfully to our family member friends and others who are directly and indirectly supported during this article.

VII. REFERENCES

1. Chandra SV, Saurabh M and Somkuwar S. Phytochemical Analysis and Antioxidant Activity of *Caesulia axillaris* Roxb. An Endemic Medicinal Plant of Indian Subcontinent. 2022; 52 (4): 112-124.
2. Farooqui M, Pradhan V, Khan J V and Khan T A. Primary phytochemical study of *Psoralea corylifolia* L. and *Caesulia axillaris* Roxb. Journal of Chemical and Pharmaceutical Research. 2016; 8(3):198-201.
3. Kumar N. *Caesulia axillaris* Roxb. leaf oil-a potential alternate of synthetic pesticides for protecting seeds of groundnut during storage. International Journal of Current Microbiology and Applied Sciences. 2015; 4(3): 372-383.
4. Matthew KM. William Roxburgh's plants of the coast of Coromandel: an enumeration of species. Blumea. 2004; 49: 367-405.
5. Naidu VSGR, Yaduraju NT and Gogoi AK. Weeds that Heal, National Research Centre for Weed Science, Jabalpur, India. Medicinal Asteraceae of India. 2005; Volume 1; pp 120. 39.
6. Naidu VSGR. Hand Book on Weed Identification Directorate of Weed Science Research, Jabalpur. 2012; India. Pp 354.
7. Natarajan B and Paulsen BS. An Ethnopharmacological Study from Thane District, Maharashtra, India: Traditional Knowledge Compared with Modern Biological Science. Pharmaceutical Biology. 2000; 38(2): 139-151.
8. Pradhan V, Farooqui M, Khan TA, Khan PA and Khan JV. Antimicrobial activity of *Caesulia axillaris* Roxb. and *Psoralea corylifolia*. World Journal of Pharmaceutical Research. 2017; 6(14): 352-358.
9. Saxena HO and Bramham M. The Flora of Orissa, Orissa Forest Development Corporation Ltd & Regional Research laboratory, Bhubaneswar, Odisha. 1995; 39(3): 156-170.
10. Shrivastava C and Jain A. Phenological events and medicinal importance of some weeds of family Asteraceae. Indian Journal of Applied and Pure Biology. 2014; 29(2): 277-282.
11. Sunatr I. The Medicinal Value of Asteraceae Family Plants in Terms of Wound Healing Activity. Journal of Pharmaceutical Sciences. 2014;39: 21-31.
12. Ethnomedicinal Use, Phytochemistry, and Other Potential Application of Aquatic and Semiaquatic Medicinal Plants. Evid Based Complement Alternat Med. 2014; 29(2): 28-36.
13. Garg, SN., Agrawal SK, Duhan SPS. Chemical examination of the leaf oil of *Caesulia axillaris* Roxb. J. Essent. Oil Res., 1993; 5: 139-142.

14. Patil MB, Khan PA and Shaikh M. Ethno Ecological Knowledge and Medicinal Flora from South-Western Satpuda, Maharashtra. Life Sciences Leaflet, 2015; 63: 44-56.
15. Shukla AC, Pandey KP, Mishra RK, Anupam Dikshit, Neetu Shukla. Broad spectrum antimycotic plant as a potential source of therapeutic agent. Journal of Natural Products, 2011; 4: 42-50.
16. Patil MB, Khan PA. Antibacterial Activity of Leaves of *Nyctanthus arbor-tristis* L, *Hibiscus rosa-sinensis* L. and *Sapindus emarginatus* Vahl. Science Park Research Journal, 2015; 31:1-5.
17. Sunccia Kocic-Tanackov, Grodana Dimic, Jelena levic, Ilija Tanackov and Danijela Tuco. Antifungal activities of basil (*Ocimum basilicum* L.) extract on *Fusarium* species. African Journal of Biotechnology, 2010; 10(50): 10188-10195.
18. Patil MB, Khan PA. Ethnomedicinal Studies of *Acalypha Indica* L. (Euphorbiaceae). Review of Research Journal, 2015; 4:7: 1-5.
19. Patil MB, Khan PA. Review: Techniques towards the Plant Phytochemical. International Journal of Science Info, 2016; I.3: 157-172.
20. NK Dubey, KA Singh, P. Shukla. R. Microbial contamination of raw materials: a major reason for the decline of indias global herbal market. Curr. Sci, 2008; 94-95.
21. Servili, M., B. Sordini, S. Esposito, S. Urbani, G. Veneziani, I.D. Maio, R. Selvaggini, and Taticchi. Biological activities of phenolic compounds of extra virgin olive oil. Antioxidants, 2020; 84-85.
22. Khan et al. World Journal of Pharmaceutical Research, 2014; 3(1): 1-23.
23. Sokmen A, Jones BM and Erturk M. The in vitro antibacterial activity of Turkish medicinal plants. J Ethnopharmacology, 1999; 67: 79-86.
24. Sarawat A K. and S. Chand. Continuous somatic embryogenesis and plant regeneration from hypocotyl segments of *Psoralea corylifolia* Linn. An endangered and medicinally important Fabaceae plant. Current Sci, 2011; 18(10): 1328-1331.
25. Tambe V N, Aher N B, Deshmukh D B and Dale D M. A research on phytochemical analysis of bioactive compounds in some medicinal plants of Asteraceae family. World Journal of Pharmaceutical Research. 2021;10 (14): 1076-1084.
26. Varma J, Tripathi M, Ram V, Pandey VB and Dubey NK. γ -Asarone- the fungitoxic principle of the essential oil of *Caesulia axillaris*. World Journal of Microbiology and Biotechnology, 2002;18: 277-279.
27. Asthana, A and Singh, A.K Fungitoxic properties of essential oil of *Ocimum adscendens*. Journal of Indian Botanical Society Supplement, 1981; 60.
28. Bocher, O.E Antibiotics. In Modern methods of plant analysis. Eds. Peach K and Tracey M.V(ed.). Modern methods of plant analysis vol iii, 651, Springer Verlag, Berlin, 1938; 10 (14):107-28.
29. Booth, C The genus *Fusarium*. Commonwealth Mycological Institute, Kew, Surrey, England, 1971; 12(9):237.
30. Chaturvedi, R Evaluation of higher plants for their fungitoxicity against *Helminthosporium oryzae*. Ph.D Thesis Gorakhpur University, Gorakhpur, India. 1979;18: 277-89.
31. Collins, P.J., Darglish, G.J., Pavic, H., Lambkin, T.M., Kapittke, R. Combating strong resistance to phosphine in stored grain pests in Australia. In: Wright, E.J., Banks, H.J., Highley, E. (Eds.), Stored Grain in Australia Proceedings of the Australian Postharvest Conference, Adelaide, 1 Technical 4 August. CSIRO Stored Grain Research Laboratory, Canberra, Australia. 2002; 109-112.
32. De Tempe, J The blotter method of seed health testing. Proc, Int. Seed Assocn, 1953; 28:133-151.
33. Dixit, V. Evaluation of volatile inhibitors from higher plants against storage fungi of *Allium cepa*. Phd thesis Gorakhpur University, Gorakhpur, India. 1954; 28: 133-151.
34. Dixit S.N., Tripathi, N.N and Tripathi, S.C Fungitoxicity of some seed extracts. Nat. Acad. Sci. Letters, 1978; 1: 287-288.
35. Bajaj, B.S., Ghosh, A.K., Antifungal antibiotics in perspective. In: Ray Chaudhari, S.P., Verma, A., Bhargava, K.S. Mehrotra, B.S. Eds., Advances in Mycology and Plant Pathology. Sagar Printers, New Delhi. 1975; 297-309.

36. Chandra, H. Evaluation of some higher plants for their volatile activity against blue mould rot of oranges. PhD The sis. Gorakhpur University, Gorakhpur, India. 1984; 297-309.
37. De Tempe, J. The blotter method of seed health testing. Proc. Int. Seed Test. Assoc. 1953; 21:133-151.
38. Dikshit, A., Dubey, N.K., Tripathi, N.N., Dixit, S.N., Cedrus oil a promising storage fungitoxicant. J. Stored Prod. Res. 2019; 19:159-162.
39. Dubey, N.K., Bhargava, K.S., Dixit, S.N. Protection of some stored food commodities from fungi by essential oils of *Ocimum canum* and *Citrus medica*. Int. J. Trop. Plant Dis. 1983; 1:177-179.
40. Funder, S., Practical Mycology: Manual for Identification of Fungi. Hafner Publishing, New York.1968; 9: 112-114.
41. Garcha, H.C., Singh, Y., Post harvest diseases of fruits in Panjab. Indian Phytopathol. 1980; 33: 42-47.
42. Moore, G.S., Jaciow, D.M. Mycology for the Clinical Laboratory. Reston Publishing, Reston. 1979; 13: 65-68.
43. Muskett, E. Technique for the examination of seed for the presence of seed borne fungi. Trans. Br. Mycol. Soc. 1948; 30:74-83.
44. Eilberg, J., Sukprakarn, C. Plant oils as fumigant and contact insecticides for the control of stored product insects. J. Stored Prod. Res.1997; 33: 7-15.
45. Sharma, R.C., Yadav, P.V. Effect of fungicides on Pencil liumrot and quality of chilgoza seed. Phytopathology. 1996; 49: 77-79.
46. Tiwari, T.N., Varma, J., Dubey, N.K. Pharmacological evaluation of some bioactive plant products on albino rats. Hind. Antibiot. Bull. 1998; 40: 38-41.
47. Varma, J. Evaluation of some higher plant products against biodeterioration of some stored food commodities. PhD The sis. Banaras Hindu University, Varanasi, India.1999; 98-99.
48. Varma, J., Dubey, N.K. Prospectives of botanical and microbial products as pesticides of tomorrow. Curr. Sci.1999; 76:172-179.
49. Panda A, Misra MK. Ethnomedicinal survey of some wetland plants of South Orissa and their conservation. Ind. J. Trad. Know. 2011; 10(2): 296-303.
50. Babcock, E. B. and Navashin, "The genus *Crepis* Biblog," Genet. 1930; 7: 34-36.
51. Babcock, E. B. and Navashin, "Basic chromosome number in plants with special reference to Compositae," New Phytologist.1934; 33.
52. Darlington, C. D. and Janaki Ammal, E. K Chromosome Atlas, George Allen and Unwin Ltd, 1945; 55-57.
53. Darlington, C. D. and Janaki Ammal, E. K. and Moffet, A. A. "Primary and secondary chromosome balances in *Pyrus*," Journ. of Gen., 1930; 22:129.
54. Deshpande, P. K. "Morphology of the endosperm in *Caesulia axillaris* Roxb.," Curr. ScL, 1960; 29: 56.
55. Hooker, J. D. Flora of British India, London, 1894; 3.
56. Jitendra Mohan, K. V., Girija, P. and Panikkar, A. O. N. "Chromosome numbers in some Compositae," Curr. Sci., 1962; 31: 207.
57. Kuwada, Y. "A cytological study of *Oryza sativa*," Bot. Mag. Tokyo, 1910;24.
58. Nandi, H. K. "The chromosome morphology, secondary association and origin of cultivated rice," Journ. Gen., 1936; 23: 315.
59. Madmuthu, K. M. and Subra-maniam, M. K. A haematox, lin squash for root-tips of *Dolichos lab* Linn.," Curr. ScL, 1960; 29: 482.
60. Azas, N., Laurecin, N., Delmas, F., Di Giorgio, C., Gasquet, M., Laget, M., Timon-David, P. Synergistic in vitro antimalarial activity of plant extracts used as traditional herbal remedies in Mali. Parasitology Research, 2002; 88: 165-171.
61. Boiteau, P., M' edecine Traditionnelle et Pharmacop' ee. Pr' ecis de mati` ere m' edicale malgache. ACCT, Paris, 1986; 36: 234.

62. Hout, S., Chea, A., Bun, S.-S., Elias, R., Gasquet, M., Timon-David, P., Bal ansard, G., Azas, N., Screening of selected indigenous plants of Cambodia for antiplasmodial activity. *Journal of Ethnopharmacology*, 2006; 107: 12-18.
63. Jenett-Siems, K., Mockenhaupt, F. P., Bienzle, U., Gupta, M.P., Eich, E., In vitro antiplasmodial activity of Central American medicinal plants. *Tropical Medicine and International Health*, 1999; 4: 611-615.
64. Kohler, I., Jenett-Siems, K., Siems, K., Hernandez, M.A., Ibarra Ricardo, A.B., Walter, G., Bienzle, U., Eich, E., In vitro antiplasmodial investigation of medicinal plants from El Salvador. *Zeitschrift fuer Naturforschung, C: Journal of Biosciences*, 2002; 57: 277-281.
65. Kraft, C., Jenett-Siems, K., Siems, K., Jakupovic, J., Mavi, S., Bienzle, U., Eich, E., Herbal remedies traditionally used against malaria. In vitro antiplasmodial evaluation of medicinal plants from Zimbabwe. *Phytotherapy Research*, 2003; 17: 123-128.
66. Lavergne, R., V'era, R., M'edecine Traditionnelle et Pharmacop'ee. Etudes ethnobotanique des plantes utilis'ees dans la pharmacop'ee traditionnelle `ala R'eunion. ACCT, Paris, 1989; 451-453.
67. Guenther, Ernest. The essential oils. V. 4, Individual essential oils of the plant families: Gramineae, Lauraceae, Burseraceae, Myrtaceae, Umbelliferae, and Geraniaceae. D. Van Nostrand, 1952; 23: 46-49.
68. Anonymous, Wealth of India. Publication and Information directorate, New Delhi: Council of Scientific and Industrial Research, 1999; vol 3: pp. 17-18,
69. Bhargawa, K.S., Dixit, S.N., Dubey, N.K. & Tripathi, R.D. Fungitoxic properties of *Ocimum canum*. *Journal of the Indian Botanical Society*, 1981; 60: 24-27.
70. Guenther, E. The Essential Oils. Huntington, New York: Krieger Publishing Company. 1972; 3; 67-69.
71. Moleyar, V. & Pattisapu, N. Detoxification of essential oil components (citral and menthol) by *Aspergillus niger* and *Rhizopus stolonifer*. *Journal of the Science of Food and Agriculture*, 1987; 39: 239-246.
72. Pandey, D.K., Chandra, H. & Tripathi, N.N. Volatile fungitoxic activity of some higher plants with special reference to that of *Callistemon lanceolatus* DC. *Phytopathology*, 1982; 105: 175-182.
73. Perucci, S., Manciant, F., Ciont, P.L., Flamini, G., Morelli, I. & Macchioni, G. In vitro antifungal activity of essential oils against some isolates of *Microsporum canis* and *M. gypseum*. *Planta Medica*, 1994; 60: 184-187.
74. Saxena, B.P. & Koul, O. Essential oils and insect control. In *Cultivation and Utilization of Aromatic Plants* eds. Atal, C.K. & Kapoor, B.M. New Delhi: Publication and Information Directorate (CSIR), 1982; pp. 766-775.
75. Thapa, R.K., Agarwal, S.C., Dhar, K.L. & Atal C.K. Role of physical and chemical methods in the analysis of essential oils. In *Cultivation and utilization of aromatic plants*, eds. Atal, C.K. & Kapoor, B.M. New Delhi: Publication and Information Directorate (CSIR), 1982; 67: pp. 90-101.
76. Thompson, D.P. Fungitoxic activity of essential oil components on food storage fungi. *Mycologia*, 1969; 81:151-153.
77. Tripathi, R.D., Srivastava, H.S. & Dixit, S.N. A fungitoxic principle from leaves of *Lawsonia inermis* Lam. *Experientia*, 1978; 34: 51-52.
78. Varma, J. & Dubey, N.K. Prospectives of some botanical and microbial products as pesticides of tomorrow. *Current Science*, 1999; 76(2): 178-186.
79. O. N. Devgan and M. M. Bokadia, Isolation of 2,4,5-trirrietzloxyallyl benzene from *Cue.sulin axillnris* oil. *Aust. J. Chein.*, 1968; 21: 001-3003.
80. Khan, T., Pradhan, V., Farooqui, M., Khan, T. A., Khan, P. A., & Khan, J. v. antimicrobial activity of *caesulia axillaris roxb* and *psoralea corylifolia*, Article in *World Journal of Pharmaceutical Research*, 2017; 8: 34-37.
81. Sarawat AK. and S. Chand. Continuous somatic embryogenesis and plant regeneration from hypocotyl segments of *Psoralea corylifolia* Linn. An endangered and medicinally important Fabcaceae plant. *Current Sci.*, 2011; 18(10): 1328-1331.
82. Ayyanar M., Indian medicinal plants as a source of therapeutic agents: a review. *Int. J. Medicinal Research*, 2012;1(1): 1-24.

83. WHO. World Health Organization traditional medicine strategy: 2002-2005, Geneva, 2001; 1-52.
84. Lourenço, S.C.; Moldao-Martins, M.; Alves, V.D. Antioxidants of Natural Plant Origins: From Sources to Food Industry Applications. *Molecules*, 2019; 24: 41-42.
85. Mishra D., Chaturvedi, R.V and Tripathi, S.C The fungitoxic effect of the essential oil of the herb *Nardostachys jatamansi* D.C. *Tropical Agri*, 1995; 72(1);48-52.
86. Muskett A. Technique for the examination of seeds for the presence of seed borne fungi. *Trans. Br. Mycol*, 1948; 30: 74-83.
87. Wellman, R.H Commercial development of fungicides. In; *Plant pathology Problem and Progress* Eds Holtan et al.,1908-1958.Indian University Press, Allahabad, India, 1967; 34: 57-59.
88. Arslan, R., Bektas, N., Ozturk, Y., Antinociceptive activity of methanol extract of fruits of *Capparis ovata* in mice. *Journal of Ethnopharmacology*, 2010; 131: 28-32.
89. on carrageenan-induced paw oedema and hepatic ischaemia-reperfusion in the rat. *British Journal of Nutrition*, 2003; 102: 126-133.
90. Koster, R., Anderson, M., De, B.E.J., Acetic acid for analgesic screening. *Federation Proceeding*, 1959; 18: 412-416.
91. Morikawa, K., Nonaka, M., Narahara, M., Torii, I., Kawaguchi, K., Yoshikawa, T., Kumazawa, Y., Morikawa, S., Inhibitory effect of quercetin on carrageenan induced inflammation in rats. *Life Sciences*, 2003; 74: 709-721.
92. Huang, S.S., Chiu, C.S., Chen, H.J., Lin, S.S., Hsieh, I.C., Hou, W.C., Huang, G.J., Antinociceptive activities and the mechanisms of anti-inflammation of acetic acid in mice. *Evidence Based Complementary and Alternative Medicine*, 2011; 67: 89-91.
93. Huang, G.J., Huang, S.S., Lin, S.S., Shao, Y.Y., Chen, C.C., Hou, W.C., Kuo, Y.H., Analgesic effects and the mechanisms of anti-inflammation of ergostatrien-3 beta-ol from *Antrodia camphorata* submerged whole broth in mice. *Journal of Agricultural and Food Chemistry*, 2010; 58: 7445-7452.
94. Huang, S.S., Chiu, C.S., Chen, H.J., Lin, S.S., Hsieh, I.C., Hou, W.C., Huang, G.J., Antinociceptive activities and the mechanisms of anti-inflammation of acetic acid in mice. *Evidence Based Complementary and Alternative Medicine*, 2011; 889-892.
95. Su, S., Wang, T., Duan, J. A., Zhou, W., Hua, Y. Q., Tang, Y. P., Yu, L., & Qian, D. W. Anti-inflammatory and analgesic activity of different extracts of *Commiphora myrrha*. *Journal of Ethnopharmacology*, 2011; 134(2): 251-258.
96. Erfan Uddin AbbVie, M., Mohammed Taufiqal Islam, A., Uddin Chowdhury, A., & Rahman, K. Sedative and Analgesic Activities of *Ludwigia repens*, 2012; 89: 763-766.
97. Arya AK, Durgapal M, Bachheti A, Deepti, Joshi KK, Gonfa YH, Bachheti RK, Husen A, 2022; 58: 78-82.