



INTERNATIONAL JOURNAL OF CREATIVE RESEARCH THOUGHTS (IJCRT)

An International Open Access, Peer-reviewed, Refereed Journal

IR-Eco Spectra: Exploring Sustainable Exsitu & In Sity FTIR Application For Green Chemical & Pharmaceutical Analysis.

Mr. Waghmare A.D., Mr. Dr. Kohle S.D., Miss . More pratibha .R

Student of bachelor of pharmacy, Anand Charitable Sanstha's College Of Pharmaceutical Science and Research, Ashti.

Tal. Ashti Dist. Beed.

ABSTRACT

Fourier transform infrared spectroscopy (FTIR) spectroscopy provides a unique ability to detect and characterize complex chemicals in many industries, especially in medicine and pharmaceuticals, while reducing waste and reducing the need for extensive modeling or use of hazardous chemicals to minimize environmental damage. This review demonstrates the wide range of ex situ and in situ FTIR applications for species identification, characterization, and quality monitoring. The accuracy of ex situ FTIR spectroscopy in identifying impurities, monitoring crystallization processes, and modifying dosage forms can improve product quality, safety, and performance. In addition, its many capabilities, all based on green screening standards, have contributed to drug development, drug use processes, and quality control. On the other hand, in situ FTIR spectroscopy appears to be a new tool for real-time investigation of drug reactions and processes, monitoring drug release kinetics, crystallization kinetics, and surface contact, and provides valuable information. The combination of ex situ FTIR accuracy and in situ FTIR dynamic capability provides a comprehensive guide to green practices, quality control, and innovation in the pharmaceutical and medical industry. This review describes the widespread use of ex situ and in situ FTIR spectroscopy as a green medicinal chemistry tool in the chemical, pharmaceutical, and pharmaceutical industries. However, more research is needed to unlock the full potential of FTIR and develop new applications that will enhance sustainability in these areas.

KEYWORD : Finished Product; Purity; Pharmacopeias; Raw Material; Quality Control

1. INTRODUCTION

Fourier transform infrared spectroscopy (FTIR) is a powerful analytical technique used to identify and measure the chemical composition of materials. Infrared spectroscopy (IR) is based primarily on the detection of molecular vibrations excited by infrared radiation. It is based on the Fourier transform, a mathematical method that separates a signal into its frequency components. FTIR works by measuring the absorption of infrared radiation by a sample and provides information about the molecular vibrations and functional groups present in the substance, represented by a specific spectrum. This information can be used to analyze the chemical composition of the sample, determine the purity of the product, and even detect impurities. The transmitted light can be controlled to ensure that energy is absorbed at each wavelength. By analyzing the powder absorption and comparing it with the chemical spectrum, information can be obtained that will allow the identification of the compound. The FTIR measurement principle is shown in Figure 1. This IRE acts as a waveguide for infrared light and as the light propagates through the sample, it interacts with the chemical functional group of the sample, creating an absorption band that can be used to control sample elements. Samples can be measured in wet or dry conditions. It is best to dry the sample before testing to reduce moisture and therefore OH interference in the IR spectrum. However, ATR allows the measurement of samples in liquid, solid and gaseous materials [1,2]. Infrared spectroscopy is a fast technology recognized for its analysis due to its many advantages such as green technology, low environmental impact since no organic solvent is used and leading the industry by reducing pollution. This even eliminates waste materials from the routine process. Spectroscopy in the infrared range is considered a simple test that can distinguish compounds with small changes and is one of the most widely used methods for cation identification [3, 4]. To maintain accuracy and efficiency, FTIR spectroscopy should be performed with appropriate spectral parameters. This is done immediately after data collection to eliminate or reduce unwanted signals in the spectrum. Misuse of these methods can compromise data reliability [5]. Techniques such as Raman spectroscopy and highperformance liquid chromatography (HPLC) can achieve this, but when combined with FTIR, they offer a unique comparison of applied and green chemistry contents. Raman spectroscopy is frequently used in medicine and chemical analysis and has significant disadvantages compared to infrared spectroscopy. Although Raman provides useful information, it requires high standards and a homogeneous reaction mixture. Fluorescence can easily interfere with Raman instruments, so it is necessary to know the initial features and characteristics in the Raman spectra. HPLC analysis provides the highest level of sensitivity for the separation and quantification of compounds. However, disadvantages such as greater solvent usage, longer processing times, higher costs, more production waste and again more labor prevent this benefit. Therefore, considering the general benefits of FTIR spectroscopy with green chemical elements and its benefits in rapid research, less preparation is required for the identification of drugs; we focus on these specific procedures in our review.[1]

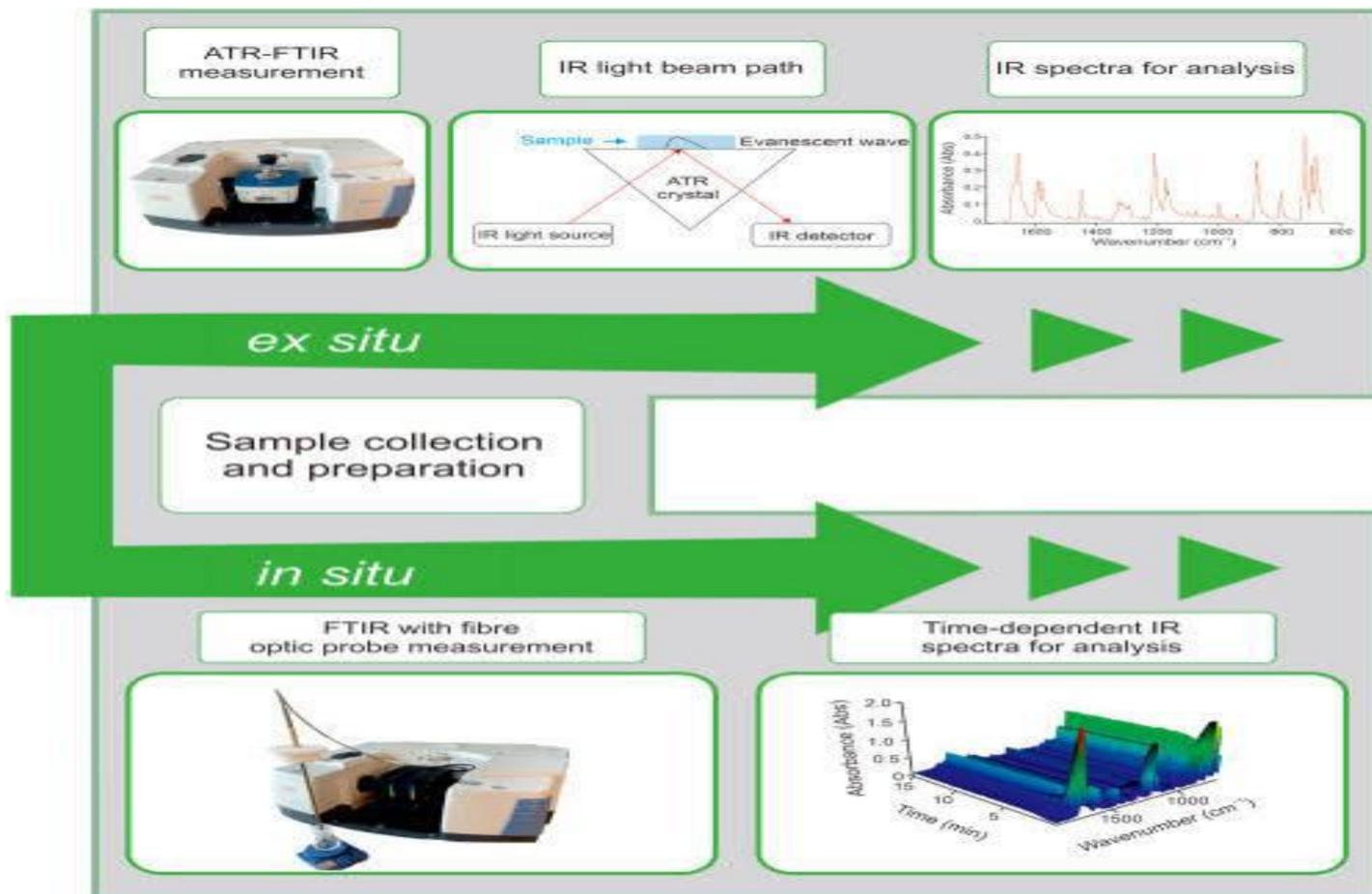


Fig. 1. The mechanism of ex situ and in situ attenuated total reflection-Fourier transform infrared spectroscopy (ATR-FTIR) use for analysis of various samples. IR: infrared.

2. Brief history of FTIR

The history of FTIR spectroscopy dates back to the 1800s when William Herschel discovered infrared radiation and later developed infrared spectroscopy [2]. Dispersive infrared spectrometers, developed in the 1940s, use prisms or gratings to disperse infrared light and measure its absorbance [3]. In the 1950s, FTIR spectroscopy technology was developed using interferometers to detect infrared light interference and provide spectral information [4]. Arthur Schawlow and Charles Townes pioneered the FTIR spectrometer in 1960 and received the Nobel Prize in Physics in 1981 for their work on laser spectroscopy. The first commercial FTIR spectrometers were introduced in the 1970s and became popular in the petrochemical and polymer fields for the analysis of organic molecules [8,9]. FTIR became a method for identifying unknown substances in drug discovery and analysis in the 1980s [10]. The invention of micro-FTIR also allows infrared spectroscopic analysis of small samples such as single cells and bacteria [5]. The introduction of portable and handheld FTIR equipment in the 1990s led onsite inspections in areas such as environmental monitoring and plant management. In the 2000s, advances in FTIR imaging technology made it possible to visualize the molecular and structural distribution of samples, and samples began to be used in disciplines such as biological sciences and information science [6]. It has been used to study everything from the molecular structure of proteins and DNA to the composition of complex polymers and drugs. Therefore, FTIR has become an important tool for scientists and researchers in many fields, including chemistry, biology and information science, in recent years. FTIR as a green and sustainable chemistry tool

The terms "green chemistry" and "sustainable chemistry" are often used interchangeably, which can lead to misunderstandings about the different meanings of the two fields. Green chemistry focuses on designing and optimizing chemical processes to minimize harmful products. In contrast, sustainable chemistry encompasses a broad framework that integrates economic and social considerations in relation to a holistic approach to service efficiency and environmental stewardship. Although green and sustainable chemistry both aim to reduce environmental impacts and promote good chemical practices, they operate in different ways [7]. The potential for confusion highlights the importance of clearly interpreting and clarifying these terms to clarify the discussion on health. It involves the design, development, and use of chemical processes and products to reduce or eliminate the use and production of hazardous substances, as well as to reduce or eliminate waste and conserve electricity and resources. Green chemistry aims to promote long-term expansion by reducing the environmental impact of chemical products and processes while maintaining their economic and operational efficiency [8]. Green chemistry is important because it reduces the adverse effects of molecular diseases and hazards from products and reagents to humans and the environment by increasing safety and chemical activity activity. Anastas and Warner proposed 12 principles of green chemistry in 1998. Green analytical chemistry emerged around 1995 as a branch of green chemistry that focuses on how analytical chemists can make their experiments more environmentally friendly. It caught the attention of pharmacists: in addition to the development of tools and methods to improve the quality of drug testing, there is an increasing effort to reduce the negative impact of drug testing on the environment and to use sustainable development standards in laboratories. Green analytical chemistry should be seen as a driver of progress in analytical chemistry. The biggest challenge for the future in this field is to find the balance between improving the quality of results and making the screening process more environmentally friendly. Sustainable Chemistry aims to increase the efficiency of the production of natural products to meet human needs for pharmaceutical products and services, while maintaining and developing environmental quality and call [9]-3 3 3

3 FTIR processing green technology and sustainable chemistry

The terms "Green Chemistry" "Sustainable Chemistry" "" and "sustainable chemistry" are often used interchangeably, which can lead to a misunderstanding of the difference in meaning between the two businesses. Green chemistry focuses on designing and optimizing chemical processes to minimize harmful products. In contrast, sustainable chemistry encompasses a broad framework that integrates economic and social considerations in a holistic approach to service efficiency and environmental responsibility. Although both seek to reduce environmental impact (FTIR), the terms "green chemistry" and "sustainable chemistry" are often used interchangeably as a form of green and sustainable chemistry, which can lead to the two misunderstandings and different meanings. Green chemistry focuses on designing and optimizing chemical processes to minimize harmful products. In contrast, sustainable chemistry encompasses a broad framework that integrates economic and social considerations in a holistic approach to service efficiency and environmental responsibility. Although both aim to reduce environmental impact and promote chemical, green and sustainable chemistry serve different purposes [10]. The like-lihood of confusion highlights the importance of precise definitions and clarification of these concepts to ensure accurate discourse within the field of science. Green chemistry is an approach to designing and developing chemical products and processes that are ecologically sustainable, cost-effective, and safe for human health. It entails the design, development, and implementation of chemical processes and products that reduce or eliminate the use and creation of hazardous compounds, as well as the reduction or elimination of waste and the conservation of energy and resources. Green chemistry seeks to support long-term expansion by decreasing the environmental effects of chemical products and processes while preserving their economic feasibility and performance [18]. Green chemistry is crucial because it reduces molecular pollution and the negative effects of hazardous by-products and reagents on people and the environment by improving the safety and efficiency of chemical operations [11]. Anastas and Warner

introduced the 12 principles of green chemistry in 1998. However, these were primarily intended for synthetic chemistry. Green analytical chemistry emerged as a branch of green chemistry around 1995, focusing on how analytical chemists can make their laboratory practices more environmentally sustainable [It has gained considerable attention from chemists: besides improving instrumentation and methodologies to enhance the quality of chemical analyses, there is a growing effort to reduce the negative impact of chemical analyses on the environment and implement sustainable development principles in analytical laboratories. Green analytical chemistry should be acknowledged as a driving force for progress in analytical chemistry. The biggest challenge for the future of this field is finding a balance between improving the quality of results and making analytical methods more environmentally friendly. On the other hand, sustainable chemistry is a more comprehensive approach, considering sociological and economic factors in addition to the fundamentals of green chemistry. Sustainable chemistry aims to increase the effectiveness of exploiting natural resources to satisfy human demand for chemical goods and services while safeguarding and improving environmental quality and human health. Although it adheres to the principles of green chemistry, it also emphasises societal welfare and economic development. In order to break the link between economic growth and environmental destruction, sustainable chemistry encourages ethical consumption and production. It adheres to the circular economic model, which aims to extend the lifespan of materials. FTIR is recognised as a green and sustainable chemistry instrument in chemical and pharmaceutical analysis for several essential reasons. Firstly, FTIR analysis aligns with the principles of green chemistry by minimising the production of hazardous waste and decreasing the use of toxic chemicals. Reduced waste and chemical use make the analytical process safer and more ecologically friendly. Secondly, FTIR makes it possible to analyse materials in real-time, obviating the need for time-consuming sample preparation and conserving energy. By encouraging energy savings and expediting the analytical process, this function is in line with sustainable and green chemistry concepts. The expedited analytical procedure advances sustainability by reducing resource consumption and encouraging energy efficiency, which aligns with broader sustainability objectives. Additionally, the molecular structure of chemicals is effectively revealed by FTIR, which helps to create chemical and pharmaceutical procedures that are more ecologically friendly. This capability promotes green and sustainable chemistry ideals by encouraging the development of safer and more effective chemical processes that eventually contribute to a sustainable future. The alignment of FTIR with the 12 principles of green chemistry is shown in Table 1. In conclusion, FTIR is a valuable tool that embodies the principles of both green and sustainable chemistry in chemical and pharmaceutical analysis, because it can analyse samples without potentially dangerous chemicals, perform in situ analysis, and provide information for more environmentally friendly chemical processes.[13]

4. Use of ex situ FTIR spectroscopy as a green analytical tool in

Ex situ FTIR spectroscopy in the pharmaceutical and chemical industries, where samples are analyzed that are not part of the reaction or process being studied. Unlike traditional methods, samples are prepared individually and then analyzed using FTIR. Ex situ FTIR spectroscopy is used in the pharmaceutical industry for material, quality control and contamination monitoring purposes. It identifies impurities, by-products and contaminants in raw materials and finished products. In addition, it can examine properties such as crystallinity, polymer composition and surface activity, all of which are important for the creation of new materials and the use of nanotechnology. This analytical technique is also widely used in the pharmaceutical industry for drug development, safety assessment, and pharmacokinetic studies. It helps ensure product quality, safety, and efficacy by analyzing drug-exipient interactions and quantifying drug content in dosage forms. Additionally, ex situ FTIR spectroscopy can be used to identify polymorphs, degradation products, and drug release patterns, providing valuable information for drug development.[14]

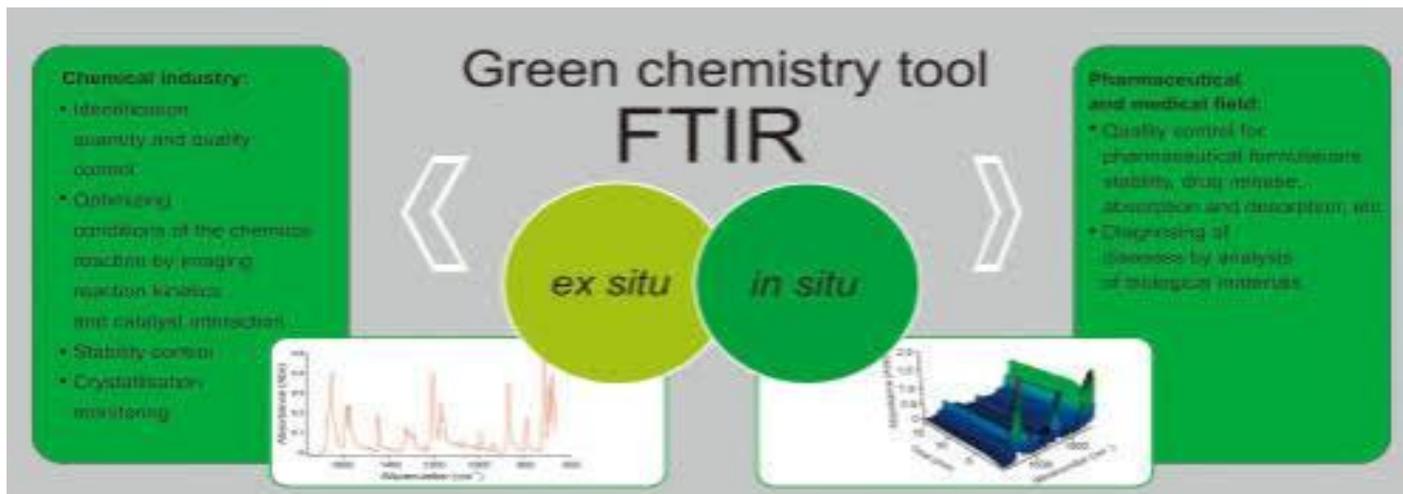


FIG 2. Green chemistry tool FTIR

Table 1 Alignment of Fourier transform infrared spectroscopy (FTIR) with the 12 principles of green chemistry.

No	Green chemistry principle	Description of FTIR usage aligning with green chemistry principle
1	Prevent waste	FTIR prevents the formation of unwanted by-products by real-time monitoring and immediate adjustments to reaction conditions and controlling the purity of substrates
2	Maximise atom economy	FTIR aids in achieving high atom economy by optimising reactions to minimise waste and maximise the conversion of starting materials into desired products.
3	Design less hazardous chemical syntheses	FTIR enables the identification and mitigation of the formation of hazardous intermediates or by-products.
4	Design safer chemicals and products	FTIR helps select reaction conditions that minimise the generation of toxic substances, as well as access the benefit-harm qualities of chemicals.
5	Use safer solvents and reaction conditions	FTIR helps choose greener solvents and auxiliaries by evaluating their interactions with reactants and products.

6	Increase energy efficiency	FTIR guides the selection of optimal reaction conditions, reducing energy consumption.
7	Use renewable feedstocks	FTIR monitors reactions involving renewable feedstocks and assesses their conversion into valuable products.
8	Avoid chemical derivatives	FTIR helps reduce the undesired derivatives by ensuring reactions proceed efficiently and by monitoring the purity of products.
9	Use catalysts, not stoichiometric reagents	FTIR assists in optimising catalyst usage by studying catalyst-reactant interactions.
10	Design chemicals and products to degrade after use	FTIR assesses the stability of products and materials, aiding in the design of substances that degrade more readily.
11	Analyse in real time to prevent pollution	FTIR's real-time monitoring helps prevent pollution by detecting and addressing issues during chemical processes.
12	Minimise the potential for accidents	FTIR provides early detection of potential hazards and enables timely interventions to prevent accidents.

The next section of the study will discuss the uses and improvements of ex situ FTIR spectroscopy in the chemical and pharmaceutical sectors, focusing on its contributions to environmentally friendly and efficient analytical practices. We aim to highlight the importance of ex situ FTIR spectroscopy as a beneficial tool in these critical areas by diving into individual case studies and significant research activities.

4.1 Identification, quality and quantity control

FTIR is widely used to determine and monitor the purity of raw materials and finished products to ensure that they meet appropriate standards. For example, FTIR technology can identify the composition of β -cyclodextrin and curcumin, since curcumin itself is a medicinal substance that requires the use of precipitation and other methods to improve stability and solubility due to its low water solubility. Another good example is the recognition obtained by Pedroso and Salgado [14], which showed that the FTIR method is suitable for the measurement of ertapenem sodium, a parenteral β -methylcarbapenem antibiotic. This method allows the characterization and quantification of ertapenem sodium without the need for organic solvents, since this sample is made in potassium bromide as a specific reagent. Therefore, this method can help reduce organic waste during mass production. Farooq et al. [27] conducted

quality control of diabetes drugs such as repaglinide, rosiglitazone maleate, pioglitazone hydrochloride and metformin hydrochloride using FTIR method and misjudged their application in quality control procedures and drug identification. Quality control is important to ensure safety and efficacy of drugs. FTIR method is a useful method for monitoring quality control of drugs and is used in identification of many drugs. The first step of quantitative investigation using FTIR is to prepare information about the known structure of the target drug to compare its chemical composition with the sample. The resulting spectrum is compared with a reference spectrum to determine the drug concentration in the sample. This can be done by direct comparison to estimate the distribution of the data or by using mathematics such as partial least squares (PLS) regression or principal component analysis (PCA). Once the relationship between the spectral data and the chemical concentration is established, the FTIR spectrum of the unknown sample can be used to determine its concentration. It is worth noting that the accuracy of FTIR quantitative analysis is affected by many factors, including the quality of the spectrum data, the stability of the sample and the application, and the resolution problem of the FTIR spectrometer. These details need to be carefully considered to ensure that the analysis results are accurate and reliable. Although somewhat selective, infrared spectroscopy provides evidence of specific functional groups, providing simplicity and cost-effectiveness compared to other methods while maintaining high accuracy [15]. The FTIR method has been used to measure ampicillin sodium in powder for injection in the range of 1.0–3.0 mg/pellet [16]. It has good linearity, precision, accuracy and robustness and is suitable for routine monitoring testing. Operation. This makes it compatible with other methods used for the same purpose such as HPLC, fluorimetry and chemiluminescence spectroscopy. Infrared has also been used to develop and evaluate methods for the measurement of ceftazidime in injectable drugs [17]. The method is based on the measurement of the absorption loop at an average of 1,475 to 1,600 cm^{-1} and exhibits good linearity, precision and accuracy over the dose range of 0.5 to 7.0 mg. Excipients do not affect the assay and the mean recovery is $98.98\% \pm 0.70\%$. Other tests using the same method include cefuroxime injection [30] (linear range 5.0–20.0 g/mL, good accuracy, standard deviation of the value is close to 2% and less than 5%), doxycycline raw material [18] (linearly in the concentration range 0.5 to 2.5 mg, correlation value 0.9991, limited test and quantification 0.125 and 0.378 mg, respectively), darunavir in tablets [19] (linearly in the concentration range 1.5e3.5 mg, correlation coefficient greater than 0.9991, detection and limit feed contain 0.12 mg and 0.36 mg, respectively) and therefore Open. Fakhlebom et al. [19] found that ATR-FTIR spectroscopy is an effective method for the measurement of diclofenac sodium in tablet formulations. Tablet excipients do not affect the linearity of the test and method with a correlation value of 0.9994 over drug amounts ranging from 0.2% to 1.5% (m/m). High recoveries (99.81%, 101.54% and 99.41%) and low detection and limiting effects reflect the high and accurate method. HPLC measurement of ketoprofen/scopolamine and benzocaine/dextromethorphan hydrobromide in binary drug mixtures and drug formulations [20]. Statistical comparisons were found to be almost the same, demonstrating its suitability for many businesses. FTIR analysis is a greener and more economical option than HPLC due to less solvent usage, mobility, reduced waste, faster processing, energy consumption. Less electricity and greater operator safety. However, HPLC analysis is quite useful in the separation and measurement of compounds, but this advantage also brings disadvantages such as additional solvent usage, longer working hours, more money spent, more waste and more effort. The specific needs of the analysis, the characteristics of the model, control limits and safety objectives should determine the choice of analytical equipment..

4.2. Crystallisation monitoring

Crystallization analysis is important in the pharmaceutical and pharmaceutical industry because it has a direct impact on product quality, purity, and efficiency. Researchers can use ex situ FTIR as an important analytical tool to analyze the crystallization process, improve efficiency, and control the industry while adhering to safety standards. Ex situ FTIR was used to study the crystallization process, which was demonstrated by analyzing the transformation of amorphous acetaminophen before and after the addition of trehalose and melibiose. Spectral analysis revealed changes in the molecular shape and interactions of the nonreactive material, including the product quenched before milling and the crystalline powder product. Amorphous materials exhibit larger and less well-defined peaks in the spectrum compared to crystalline materials [In addition to pharmaceutical applications, FTIR spectroscopy also shows potential as an analytical tool to advance protein crystallization research. This

continues to explore the influence of surface properties on protein behavior and address important questions regarding protein structure and function [36]. Additionally, FTIR technology can measure and monitor the crystallization and melting properties of various vegetable oils, as exemplified by the study of Physalis peruvica [21]

4.3. Oxidation and stability control

Oxidation processes are important in chemistry, medicine and biology, especially in the development of living organisms. Oxidation is defined as the loss of electrons from a molecule or atom, usually by the addition of oxygen or the removal of hydrogen. Oxidation is the second most common pharmacological degradation mechanism after hydrolysis and is very important for the development of new materials and synthetic drugs because it can add functional groups, change chemical structure and form valuable products. Controlling and characterizing oxidative processes involved in drug metabolism is important for drug development, drug use and prediction of drug interactions, as well as for understanding the chemical process and improving synthesis. Surapaneni et al. [22] studied the oxidation of (-)-menthol to (-)-menthone in different solvents using FTIR spectroscopy. The solvents used in this reaction (acetic acid, acetone, ethyl acetate and methylene chloride) have a significant effect on the reaction kinetics and yield. Due to the increase in hypochlorite, which is the limit of the oxidation reaction, the authors consider that the most polar solvent system will make the reaction fastest and best. The results showed that the reaction using the least amount of solvent, ethyl acetate, gave the best yield of (-)-menthone. Surprisingly, the reaction time was shortest in the solvent systems ethyl acetate and acetic acid, both of which are less polar than the primary solvent used for this reaction, acetonitrile. The authors used FTIR spectroscopy to characterize all reaction products based on (-)-menthone, which provided insights into the effects of solvent selection. This finding highlights the importance of using green chemistry techniques when selecting solvents for chemical reactions. Pharmaceutical formulations often contain a lipid phase dispersed in an aqueous medium, forming an oil-in-water (o/w) emulsion. These emulsions are important for monitoring lipid oxidation. The mechanism of O/W oxidation is more complex than that of bulk oil oxidation. Water-soluble and oil-soluble pro-oxidants and antioxidants can interact at the oil-water interface. Therefore, there is an urgent need for accurate and rapid analytical methods to directly measure the oxidation of drugs and chemicals, and FTIR has been shown to be an effective method to analyze omega-3 fatty acid oxidation, as demonstrated by the method of Daoud et al. [23]. FTIR spectroscopy, combined with recent advances in data processing and chemometrics, becomes suitable for investigating the stability and suitability of biopharmaceuticals, such as functional properties of biomaterials, monitoring of monoclonal antibody purification, and bioactivity of biopharmaceuticals under stress.

4.4. Drug dissolution and release control

The study of drug release and solubility is important in finding new drug candidates. For preclinical testing, it is recommended that the solubility of the drug be greater than 10 mM. Analysis of early preclinical data to determine solubility can help determine the need for a resource-intensive formulation. On the other hand, changes in chemical structure due to degradation and dissolution of chemical components will reduce the potency of the drug, increase performance issues, and no longer have sufficient safety. Fear that product degradation can be dangerous. The study of forced degradation is one of the first steps in the development of a new drug and provides the first insight into its medicinal properties. Researchers can use FTIR data obtained from forced degradation to evaluate drug stability at different sites based on the amount of degradation impurities produced, such as the darunavir study by Modini et al. and the doxofylline and deflazacort study by Raju et al. [24]. Abdul Halim et al. [4] attempted to use the synergistic effect of lavender essential oil to create a self-

nanoemulsified drug delivery system to improve the poor permeability and side effects of zolmitriptan (ZMT) and increase its efficacy in the treatment of migraine pain. A continuous system coupled with ATR-FTIR, water sorption, separation and computer techniques were used to estimate the percentage of dissolved ZMT over time and the permeation test. While standard analytical methods such as HPLC require continuous monitoring and re-

examination of suspended solids, FTIR analysis provides an indication of the continuous monitoring of the sequestration structure of the active substances. The modified ZMT self-nanoemulsification system produces nanoscale spheres with higher permeation power than conventional ZMT during initial dissolution. FTIR imaging has also been used to characterize the distribution of poorly water-soluble drugs in polyethylene glycol (PEG) and their separation in water [25]. It was found that amorphous nifedipine began to crystallize in PEG-8000 for formulations with at least 10% drug content (m/m). ATR-FTIR spectroscopic imaging provides new insights into the dissolution process of nifedipine from water-soluble polymer product dispersions, which is useful for optimizing formulation manufacturing. FTIR can be used to analyze the adsorption and desorption of drugs in various delivery systems. By measuring changes in the infrared spectrum, researchers can monitor the release of drugs from the formulation and understand their interactions with excipients, which is important for tuning drug delivery.

Zid et al. tested mesoporous silica as a drug delivery system for naproxen [46]. It was found that the adsorption and desorption properties of naproxen were affected by solution pH and silica surface functionality. By comparing the spectra of mesoporous silica, modified samples and naproxen-loaded materials, the grafting of organic groups onto silica and the adsorption quality of naproxen can be clearly seen.

4.5. Control of Nanoparticle Formation

Nanoparticles have made significant progress in the pharmaceutical and pharmaceutical industries. By encapsulating nanoparticles in drug-based development, researchers can improve drug stability and bioavailability and target specific cells or tissues

. This prevents drug degradation, controls their release rate and helps them pass through cellular barriers. Effectiveness [26]. Nanoparticles can also act as catalysts in many chemical processes. Due to their large area and specific nature of the area, they can realize efficient processes by reducing the need for expensive and ecologically harmful energy sources. Bashir et al. Use the green method to produce ZnO nanoparticles with large pores [27]. FTIR analysis revealed the formation of ZnO nanoparticles and the presence of phytochemicals that aid in formulation. Many studies have shown that FTIR is an important technique for the analysis of nanoparticle formation [28] and crystallinity characterization. Replication It is still an important tool due to its ease of use, flexibility and the ability to analyze many devices with similar ones. The advantage of ex situ FTIR spectroscopy as a green analysis is due to its non-destructiveness, minimal preparation and minimal waste. This approach supports sustainable practices by saving resources and providing valuable information without compromising the integrity of the analysis model.

5. Use of in situ FTIR spectroscopy as a green analytical tool in the chemical and pharmaceutical industry

In situ FTIR spectroscopy works on the same principle as traditional FTIR, where an infrared beam is transmitted through a sample and the absorbed energy produces a specific spectrum corresponding to the molecular vibrations in the structure. In traditional ex situ FTIR spectroscopy, samples are prepared separately and then analyzed in a spectrometer. This method provides important information about the importance of the structure, but cannot capture the dynamics of the entire reaction or process. In situ FTIR spectroscopy, on the other hand, requires the sample to be immediately placed in the FTIR spectrometer or the spectrometer to be connected to the reaction vessel. This allows the pattern to be continuously monitored as it changes, allowing for timely resolution of the progress of drug systems and time-

sensitive processes. The main work of the research is to show the impact of changes in the field of FTIR spectroscopy based on the use of green plants. Identification, Quality Control and More Similar to ex situ FTIR, in situ FTIR is frequently used for sample analysis, especially in commercial medical fields. It plays an important role in defining product purity, detecting impurities and identifying products resulting from the degradation process as suggested in various studies [61 and 65]. Powerful tools to ensure consistency and compliance with manufacturing standards in the pharmaceutical and pharmaceutical industries. An exemplary study by

Chan et al. involved the creation of ibuprofen formulations in PEG covering a range of ibuprofen concentration from 0% to 100%. This is done using the droplet deposition method. The ibuprofen concentration in the PEG matrix is determined by measuring all samples simultaneously using in situ FTIR spectroscopic imaging. The analysis of FTIR spectra of these samples provides information on the molecular state of the drug and the length of polymer swelling, which is affected by various chemical factors. Another good example is given by Ho et al. [29] used in situ FTIR spectroscopy to investigate the formation of fucoidan/chitosan based polyelectrolyte multilayers (PEM) and used ATR-FTIR to monitor the continuous growth. The intensity of the different peaks associated with each polymer increases during each adsorption step and spectral analysis allows the extraction of layer-specific spectra, facilitating the value of the adsorbed mass at each stage. Proton modifies membrane synthesis.

5.2. Reaction kinetics and catalyst interaction control

Small differences in reaction kinetics are important for developing effective processes and good products in drug and pharmaceutical research. A good understanding of reactions, mechanisms, catalyst effects and reaction parameters is important. In this context, in situ FTIR seems to be an important tool that allows researchers to grasp the complexity of reaction kinetics and make informed decisions about catalysts and reaction decisions to achieve the best. In situ FTIR

provides dynamic information about the dynamics by observing the stable molecular vibrations. To calculate the reaction kinetics, data on concentration versus time should be collected. Beer's law states that the absorption of a substance in a mixture varies with the concentration of that substance. As reactants are converted to products, their characteristic infrared absorption bands change, getting stronger or weaker. These spectral changes can be recorded in situ by FTIR, providing detailed information about the outcome of the event. By calculating the rate of spectral changes, scientists can identify reactions, find rate-limiting processes, and even find intermediates that escape scientific formulas. The ability to follow reaction kinetics at the molecular level allows researchers to accurately target negative reactions [30]. Selectivity, yield and reaction method are all strongly affected by the catalyst used. In situ FTIR provides a unique advantage in measuring the interactions between reactants and catalyst surfaces. Changes in the infrared spectrum indicate the adsorption process, the type of association and the active sites where reactant molecules adsorb onto the catalyst. This information is important for understanding how various catalysts affect the reaction process and has been used in many studies over the past few years. Deactivation, regeneration and ideal operating environment.

According to research by Marinkovic et al. [79] used the electrochemical in situ FTIR technique to demonstrate the role of Rh in promoting CeC bond cleavage in ternary PtRh/SnO₂ catalysts. In addition, in situ FTIR provides quantitative confirmation of the increase in the overall oxidation process leading to CO₂ production. This can be done by in situ FTIR acting as a reporter. Using these tools, scientists can quickly assess how these changes affect reactions, equilibria, and explosions. This rapid feedback facilitates finding the best sites for reactions that produce better products.

5.3. Drug release control

Appropriate control of drug release from drug formulations is an important aspect of current drug delivery aiming to increase efficacy while minimizing side effects, and in situ FTIR has proven itself as a powerful tool. Furthermore, FTIR imaging helps in visualizing changes in drug release over time. Unlike traditional degradation studies, this spectral imaging technique provides new insights into the changes that occur during drug degradation. In situ FTIR imaging shows great potential in various applications including studies on multilayered tablets, simplified high-

throughput studies, use of microfluidic devices and in situ surface treatment ATR - FTIR spectroscopy developed a combined ATR-

FTIR spectral imaging method and designed a polydimethylsiloxane microfluidic device specifically to study drug release from drug formulations. The separation of microformulations such as ibuprofen and PEG in flowing water was investigated and the behavior and release of the drug in different pH environments were observed. It can be seen from the spectral image and ATR-

FTIR spectrum that the drug transforms from the molecular state to the crystalline form in acidic solution. Mi

crofluidic devices allow multiple microformulations to be exposed to different liquids simultaneously and provide an efficient way to study multiple microformulations in a single experiment. This study also highlights the importance of analyzing the behavior of the drug after release (such as recrystallization from solution). A TR-

FTIR spectral imaging data show the phase change of ibuprofen sodium dissolved in a neutral solution into an acidic environment.

Change. The combination of ATRFTIR spectral replication and microfluidic devices provides a method with potential for further development, from pressure measurement to the study of drug release and product behavior in water, as a tool for learning various design and problem-solving and dynamic methods. In situ FTIR spectroscopic imaging was used with a flow cell device to study the effect of different excipients on drug release formulations.

It was found that the difference in drug release was mainly due to drug-polymer interactions, and the addition of sodium carbonate improved the release by reducing these interactions and promoting the production of more water-soluble ibuprofen.

Other successful examples of the use of in situ FTIR in drug dissolution imaging include the study of antiviral drug release by Ewing et al. [31]

5.4. Crystallisation control

The ability of a molecule to take on different crystalline forms in its components is called polymorphism. Polymorphs differ in physical and chemical properties such as lattice strength, melting point, heat of fusion, solubility, dissolution rate, speed, and processing properties. These differences can affect the stability, formulation, potency, bioavailability, storage, and efficacy of the drug. In terms of intellectual property, a particular polymorph can be patented if it exhibits better properties than a previously patented polymorph, allowing competitors to legally offer products with different crystal structures. Modification of the crystallization medium, such as changing the temperature, type of solvent, and chemical pH, can produce a variety of polymorphic forms [The polymorphic process can be closely monitored using in situ FTIR, allowing for instantaneous observation of molecular changes during crystallization. The importance of the major transition stages. Understanding the detailed metamorphism of amorphous calcium carbonate into concrete at the molecular level is critical for the removal of many biologically active compounds. In their study, the researchers used in situ FTIR spectroscopy to study the moisture-

induced crystallization process of amorphous calcium carbonate. This technique is useful for changes in the vibrational properties of carbonate and water molecules, making it a useful analytical tool. Another good example is provided by Chan et al. [66] described the use of in situ FTIR imaging in combination with a controlled humidity unit to monitor the crystallization of a binary mixture of two substances at the same site. Using an infrared focal plane array detector, the researchers studied the effect of humidity on a binary mixture of nifedipine and nitrendipine as well as amorphous nitrendipine arranged in different molar ratios on the surface of a BaF₂ window. This study also investigated the effect of sample thickness on the analysis of the results of the images using a special method for creating thickness

simple images. The FTIR spectral imaging method proposed in this paper can be used for future high-throughput studies of large numbers of samples in controlled areas. The method also demonstrates the ability to simultaneously study changes in the crystallization behavior and polymorphism of different chemicals and their compounds. The advantage of this research proposal is that each sample can be measured according to the chemical image, which allows the spectra of different polymorphs to be acquired and measured separately without requiring spectral subtraction. There are other important research examples where in situ FTIR is used to control crystallization in the pharmaceutical industry, notably the polymorphic changes of carbamazepine [87], rifampicin [88] and celerycoxib [32] and clinical trials

5.5. Absorption and desorption control

In situ FTIR spectroscopy has proven to be an effective method for studying interactions between particles. This technique is used in many scientific studies to help scientists unravel the complex dynamics of molecular adsorption and desorption processes. In situ FTIR can provide insight into the small-scale interactions between adsorbate molecules and the substrate, and in many cases can lead to a better understanding of material behavior. This technology is used in many fields, including catalysis, materials science, and environmental protection. The special technique in this process requires carefully placing a high-refractive index crystal at a specific angle to the surface pattern, thereby creating a transient wave that penetrates the material. This method allows for instant, non-destructive examination of surface layers without the need for sample preparation. Many studies have used HATR-

FTIR to study adsorption and desorption phenomena in depth. For example, a recent study investigated the desorption of glyphosate from goethite due to phosphate adsorption. Combining HATR-FTIR spectroscopy and adsorption isotherm analysis, the researchers determined that glyphosate desorption on the upper surface of the phosphate was significant. This effect occurs because the ratio of desorbed glyphosate to adsorbed phosphate is 0.60. Similarly, studies have investigated the effect of humic acid on the adsorption/desorption behavior of glyphosate on goethite and the adsorption of dimethyl sulfide on silver-modified bentonite [

Iron-

(hydroxy)dimethylarsinic acid is the main intermediate in the electrochemical reduction of CuO nanoparticles [32] and CO₂. Local patterns of chemical and different processes of water are investigated. In situ infrared spectroscopy is also an excellent tool for the quality and quantity of adsorbed water due to its sensitivity to water and hydrogen bonding states. Redistribution Fourier Transform Spectroscopy (DRIFTS) is another useful tool for studying the environment. In contrast to HATR-

FTIR, which uses a high refractive index crystal in direct contact with the sample surface, in situ DRIFTS uses a different concept. Infrared radiation should be used to illuminate the dispersion or powder sample and capture the scattered reflected light for analysis. Surface chemistry, studies have shown that Cu/Ti(H₂) exhibits more than 50% higher CO₂ photoreduction activity compared to Cu/Ti (air), which is attributed to Cu²⁺, OH. The effect of the group and oxygen-free sites thus improves energy transfer, provides CO₂ adsorption sites and promotes CO₂ activation [100]. It seems to be a powerful technique to instantly probe surface intermediates. These studies exemplify the widespread use of in situ FTIR, particularly HATR-

FTIR and DRIFTS, to investigate the interactions between surfaces and molecular dynamics. In situ FTIR has provided precise characteristics of changes in material properties, increasing our understanding of the materials used and enabling new developments in various fields such as environmental science and materials engineering. Compared to ex situ methods that require samples to be removed from their original locations, in situ FTIR can monitor molecular species adsorbed onto the material during chemical reactions. This feature is very useful because it allows the study of countermeasures, the identification of transient intermediates, and the monitoring of animal dynamics under real reaction conditions. In situ FTIR has the advantage of capturing precise details of molecular interactions and changes without affecting physical properties. This real-time analysis of surface[40]-

environment behavior allows for better insight into the catalytic process, the reaction process, and the role of many types of surfaces, ultimately paving the way for catalyst material design and optimization. This allows us to understand complex reaction mechanisms and intermediates, but the process requires special equipment and experiments can be complex, causing problems in applying models and settings.

6. Combination of FTIR technique with other analytical

Methods for optimizing chemical and pharmaceutical processes

Methodological synergy in chemical analysis involves integrating and combining different analytical methods or methods to obtain complete and reliable datasets on analysis models. Researchers can benefit from it by considering the limitations of each

strategy using different methods. This method allows users to better understand the composition, structure, properties and behavior of the model. FTIR can also be used in conjunction with other analytical methods to optimize laboratory and industrial processes. It can be combined with FTIR to observe the composition of gaseous and liquid phases during a reaction [33]. For example, Ke et al. The efficiency and reliability of the self-optimizing system were validated by two interventions, including oil-

liquid separation. a. It can be used together with FTIR to provide additional information on drug mixtures and drug formulations. As a practical example, Raman and FTIR spectroscopy were well combined with gas chromatography when optimizing the ultrasound-

assisted extraction of bioactive compounds from acacia gum using field methodology. Polysaccharides such as galactose and glucose and proteins such as lysine and proline were detected, while the FTIR spectrum showed the presence of functional groups such as alkanes, aldehydes, fatty amines and phenols. The presence of antioxidant compounds D-galactopyranose, carotenoids and lycopene was analyzed. This technique can help to identify the most significant changes occurring during the reaction and hence to optimize the reaction [39].

The combination of FTIR analysis and chemical chemometric analysis is a promising analytical tool that aims to reduce the complexity of drug analysis with green performance. Compared to traditional methods such as HPLC, it can help in reducing measurement, reducing problems, reducing work time and mobility, reducing costs, reducing waste and improving user safety. Side-by-side comparison of the potential of FTIR and HPLC techniques combined with PLS regression for quantitative identification of active compounds by green chemical analysis

Summary of the study by Kelani et al. [34].

The analysis of ketoprofen/scopolamine and benzocaine/dextromethorphan hydrobromide in binary mixtures and drug formulations using FTIR-

PLS regression shows superior performance as it provides less weight consumption, portability, less waste production, short working time, reduces operating costs,

lower energy consumption, improved operator safety and easy integration with traditional chemical equipment here. Antidote. This technique has several advantages, including the ability to control unfavorable reactions, improve mass conversion, improve reaction kinetics, and provide information about the behavior of the electrochemical reaction of reactants. Overall, combining FTIR with other techniques provides an effective method for monitoring and optimizing reactions.

7. Use of FTIR spectroscopy in the medical industry for

Diseases and Treatments The development of diseases such as cancer and infectious diseases and the lack of appropriate, reliable, cost-effective and technology-

based high diagnostics necessitate the development of other diagnostic tests. FTIR spectroscopy is important for understanding many applications from chemical and quality control to biomedicine. This method is a rapid, non-invasive, label-free, reagent-free and well-

reproducible method for the characterization of biomolecules

[It allows automated and reproducible analyses leading to objective analysis of samples. FTIR can reveal molecular structures and chemical elements such as proteins, lipids, nucleic acids and carbohydrates. It can also identify changes in the molecular composition associated with the disease by providing specific markers of biological materials such as tissues, cells and fluids. The role of FTIR spectroscopy in the diagnosis of infectious diseases [1] is studied by spectral changes, thus providing precise information about the stage of the disease. The study of genomic changes during the infection, known as MTP, will be important in the diagnosis of the disease. FTIR spectroscopy can identify changes such as total cholesterol and immunoglobulins in serum after infection. Some studies have tried to develop new diagnostic methods for the 2019 coronavirus (COVID-19) based on saliva and plasma vibration modes examined by ATR-

FTIR spectroscopy. They determined the biological fingerprints of COVID-19 and followed the diagnosis of COVID-19 using a multivariate linear regression model, which will help to develop diagnostic equipment faster and cheaper in the future. Lee-Montiel and colleagues investigated the effectiveness of FTIR spectroscopy in identifying infectious diseases [126], using poliovirus (PV1) and buffalo green kidney cells. The results showed that the method worked best at 8 h post-infection and could identify virus titers ranging from 10 to 106 PFU/mL. According to the study, this method for identifying poliovirus and its composition could be applied to other diseases and adapted for use in water safety monitoring and pain testing. FTIR studies showed that, from an optical perspective, the intensity of carbohydrate peaks decreased with the growth of herpesvirus [35]. Decreases in cellular glucose content are used as biomarkers of herpesvirus kinetics and can be used to develop spectroscopic methods to measure herpesvirus growth. ATR-FTIR spectroscopy has also been used to identify hepatitis C and B viruses caused by dengue arboviruses and dengue fever. It is a tool for the detection and analysis of different types of cell death in leukemia. The results show that FTIR spectroscopy can distinguish between apoptosis and necrosis based on changes in DNA conformation and protein secondary structure. The B-pattern is increased in association with apoptosis, while random coil formation is reduced during necrosis. Studies show that FTIR spectroscopy appears to be a promising method for in situ monitoring of cell death, as it can provide unbiased biochemical information with minimal sample processing and without the need for reagents. There are reports on the use of FTIR spectroscopy to detect cancers: breast cancer, lung cancer, cancer [36], etc. It measures urine such as protein, creatinine and urea. FTIR spectroscopy, which provides information about bone composition and molecular numbers, can be used to diagnose bone-related diseases. Using FTIR spectroscopy, researchers determined the contribution of minerals and collagen to the risk of stress fracture (SF) and determined the differences in biochemical markers between bone health and stress. According to comparative analysis and biochemical profiling, stress fractures show higher collagen content, poor growth, texture, carbonate and acid alteration, and higher Crystallinity compared to healthy bone.[38]

8. Conclusion

This review aimed to provide strong evidence of FTIR spectroscopy as a practical, fast and selective technique, with the advantage of requiring small samples, having a viable budget in terms of instrumentation, increasing the ability to identify or characterise complex structures, minimizing the handling of toxic materials, and reducing the generation of organic waste solvents. The primary contribution of this work is to show examples of ex situ and in situ FTIR applications for substance identification and analysis that can now also be utilised in chemical, pharmaceutical and medical fields, while being “greener” than existing procedures. FTIR spectroscopy may be conveniently employed in routine drug and chemical testing and quantification. Moreover, this valuable technique may be employed in disease identification.

In all industries, ex situ FTIR spectroscopy is a reliable tool for material identification, quality control, and stability assessment. Its ability to characterise contaminants, monitor crystallisation processes, and manage medication release patterns highlights its critical role in product quality, safety, and efficacy. Furthermore, its quantification capabilities lead to more efficient drug development, dosing methodologies, and quality control practices, while its environmentally friendly characteristics are consistent with green

analytical practices. In situ FTIR spectroscopy, on the other hand, emerges as a novel tool for dynamic analysis, allowing for real-time monitoring of molecular changes during reactions and processes. Its contributions range from monitoring drug release patterns and crystallisation dynamics to studying surface interactions and adsorption phenomena. The time-resolved examination of chemical reactions by in situ FTIR offers a unique perspective that complements ex situ approaches, leading to a better knowledge of molecular changes and material behaviour. Together, the precision of ex situ FTIR in characterising and quantifying, combined with the ability of in situ FTIR to capture dynamic changes, creates a comprehensive analytical framework for advancing green practices, quality control and innovation in both chemical and pharmaceutical sectors. With

their distinct advantages, these methodologies work in tandem to shape efficient, sustainable and effective analytical strategies for the growth of research and industry. Overall, the prospects for using FTIR in greening chemical and pharmaceutical processes are promising. However, further research is needed to explore the potential of FTIR in these fields fully and to develop new applications that can further enhance the sustainability of chemical and pharmaceutical processes.

Declaration of competing interest

The authors declare that there are no conflicts of interest.

Acknowledgments

This work was supported by the Large Research Grant from the Doctoral School, funded by statutory funds from Poznan University of Medical Sciences, Poland (Grant No.: 85/2023)

REFERENCE

- [1] M.C.D. Santos, C.L.M. Morais, K.M.G. Lima, ATR-FTIR spectroscopy for virus identification: A powerful alternative, *Biomed. Spectrosc. Imag.* 9 (2021) 103e118.
- [2] V. Tucureanu, A. Matei, A.M. Avram, FTIR spectroscopy for carbon family study, *Crit. Rev. Anal. Chem.* 46 (2016) 502e520.
- [3] T.M. Pedroso, H.R.N. Salgado, Green alternative using infrared spectroscopy as an efficient and stable analytical method for quantifying eritapenem sodium, *Adv. Anal. Pharm. Chem.* 2018 (2018), AAPC-103.
- [4] S.M. Abd El-Halim, M.A. Mamdouh, S.M. Eid, et al., The potential synergistic activity of zolmitriptan combined in new self-nanoemulsifying drug delivery systems: ATR-FTIR real-time fast dissolution monitoring and pharmacodynamic assessment, *Int. J. Nanomed.* 16 (2021) 6395e6412.
- [5] H. Tiernan, B. Byrne, S.G. Kazarian, ATR-FTIR spectroscopy and spectroscopic imaging for the analysis of biopharmaceuticals, *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 241 (2020), 118636.
- [6] E.F.J. Ring, The discovery of infrared radiation in 1800, *The Imaging Science Journal* 48 (2000) 1e8.
- [7] E.D. Becker, T.C. Farrar, Fourier transform spectroscopy, *Science* 178 (1972) 361e368.
- [8] J.L. Arrondo, F.M. Goni, J.M. Macarulla, Infrared spectroscopy of phosphatidylcholines in aqueous suspension. A study of the phosphate group vibrations, *Biochim. Biophys. Acta* 794 (1984) 165e168.
- [9] J.L. Koenig, M.K. Antoon, Recent applications of FT-IR spectroscopy to polymer systems, *Appl. Opt.* 17 (1978) 1374e1385.
- [10] K.S. Kalasinsky, B. Levine, M.L. Smith, et al., Detection of amphetamine and methamphetamine in urine by gas chromatography/Fourier transform infrared (GC/FTIR) spectroscopy, *J. Anal. Toxicol.* 17 (1993) 359e364.
- [11] K. Bagley, G. Dollinger, L. Eisenstein, et al., Fourier transform infrared difference spectroscopy of bacteriorhodopsin and its photoproducts, *Proc. Natl. Acad. Sci. U. S. A.* 79 (1982) 4972e4976.
- [12] J. Breton, J.R. Burie, C. Berthomieu, et al., The binding sites of quinones in photosynthetic bacterial reaction centers investigated by light-induced FTIR difference spectroscopy: Assignment of the QA vibrations in *Rhodobacter sphaeroides* using ¹⁸O- or ¹³C-labeled ubiquinone and vitamin K1, *Biochemistry* 33 (1994) 4953e4965.
- [13] M. Jackson, H.H. Mantsch, The use and misuse of FTIR spectroscopy in the determination of protein structure, *Crit. Rev. Biochem. Mol. Biol.* 30 (1995) 95e120.
- [14] K.K. Chittur, FTIR/ATR for protein adsorption to biomaterial surfaces, *Bio-Materials* 19 (1998) 357e369.

- [15] J.F. Neault, M. Naoui, H.A. Tajmir-Riahi, DNA-drug interaction. The effects of Vitamin C on the solution structure of calf-thymus DNA studied by FTIR and Laser Raman difference spectroscopy, *J. Biomol. Struct. Dyn.* 13 (1995) 387e397.
- [16] T. Noguchi, Light-induced FTIR difference spectroscopy as a powerful tool Toward understanding the molecular mechanism of photosynthetic oxygen evolution, *Photosynth. Res.* 91 (2007) 59e69.
- [17] LZid, V. Zelenak, M. Alm ————— a—————si, et al., Mesoporous silica as a drug delivery System for naproxen: Influence of surface functionalization, *Molecules* 25(2020), 4722.
- [18] M.J. Mitchell, M.M. Billingsley, R.M. Haley, et al., Engineering precision Nanoparticles for drug delivery, *Nat. Rev. Drug Discov.* 20 (2021) 101e124.
- [19] A.A. Yetisgin, S. Cetinel, M. Zuvun, et al., Therapeutic nanoparticles and their Targeted delivery applications, *Molecules* 25 (2020), 2193.
- [20] Y. Huang, J. Ren, X. Qu, Nanozymes: Classification, catalytic mechanisms, Activity regulation, and applications, *Chem. Rev.* 119 (2019) 4357e4412.
- [21] Q. Li, L. Ji, B. Jiang, et al., Pillararene-functionalized rhodium nanoparticles for Efficient catalytic reduction and photothermal sterilization, *Chem. Commun.(Camb)* 58 (2022) 13079e13082.
- [22] M. Bashir, F. Majid, R. Sabir, et al., Facile green synthesis, analysis, in vitro Antidiabetic and antimicrobial activity of ZnO macropores, *Bioprocess BioSyst. Eng.* 45 (2022) 1993e2006.
- [23] N.I. Farkas, L. Marincas, R. Barabas, et al., Preparation and characterization of ————— Doxycycline-loaded electrospun PLA/HAP nanofibers as a drug delivery System, *Materials* 15 (2022), 2105.
- [24] N. Joshi, N. Jain, A. Pathak, et al., Biosynthesis of silver nanoparticles using Carissa carandas berries and its potential antibacterial activities, *J. Sol Gel Sci. Technol.* 86 (2018) 682e689.
- [25] N.A. Begum, S. Mondal, S. Basu, et al., Biogenic synthesis of Au and Ag Nanoparticles using aqueous solutions of Black Tea leaf extracts, *Colloids Surf. B Biointerfaces* 71 (2009) 113e118.
- [26] N.I. Abdullah, M.B. Ahmad, K. Shameli, Biosynthesis of silver nanoparticles Using Artocarpus elasticus stem bark extract, *Chem. Cent. J.* 9 (2015), 61.
- [27] J. Karimi, S. Mohsenzadeh, Rapid, green, and eco-friendly biosynthesis of Copper nanoparticles using flower extract of Aloe vera, *Synth. React. Inorg. Met. Org. Nano Met. Chem.* 45 (2015) 895e898.
- [28] V. Soni, P. Raizada, P. Singh, et al., Sustainable and green trends in using plant Extracts for the synthesis of biogenic metal nanoparticles toward environmental and pharmaceutical advances: A review, *Environ. Res.* 202 (2021), 111622.
- [29] A.M. Croitoru, A. Moroşan, B. Tihauan, et al., Novel graphene oxide/quercetin And graphene oxide/juglone nanostructured platforms as effective drug deLivery systems with biomedical applications, *Nanomaterials* 12 (2022), 1943.
- [30] T.A. Saleh, Characterization and description of adsorbents and nanoMaterials, in: *Interface Science and Technology* 34, Elsevier, Amsterdam, 2022, pp. 199e232.

- [31] H. Issa. Hamoud, L. Wolski, I. Pankin, et al., In situ and operando spectroscopies in photocatalysis: Powerful techniques for a better understanding of the performance and the reaction mechanism, *Top. Curr. Chem. (Cham)* 380(2022), 37.
- [32] Y. He, F. Guo, K.R. Yang, et al., In situ identification of reaction intermediates and mechanistic understandings of methane oxidation over hematite: A combined experimental and theoretical study, *J. Am. Chem. Soc.* 142 (2020)17119e17130.
- [33] K.S. Reddy, B. Siva, S.D. Reddy, et al., In situ FTIR spectroscopic monitoring of the formation of the arene diazonium salts and its applications to the heck-Matsuda reaction, *Molecules* 25 (2020), 2199.
- [34] V. Zholobenko, F. Rutten, A. Zholobenko, et al., In situ spectroscopic identification of the six types of asbestos, *J. Hazard. Mater.* 403 (2021), 123951.
- [35] T.T.M. Ho, K.E. Bremmell, M. Krasowska, et al., In situ ATR FTIR spectroscopic study of the formation and hydration of a fucoidan/chitosan polyelectrolyte multilayer, *Langmuir* 31 (2015) 11249e11259.
- [36] H. Cheng, S. Wu, J. Huang, et al., Direct evidence from in situ FTIR spectroscopy that o-quinonemethide is a key intermediate during the pyrolysis of guaiacol, *Anal. Bioanal. Chem.* 409 (2017) 2531e2537.
- [37] K.L.A. Chan, S.G. Kazarian, D. Vassou, et al., In situ high-throughput study of drug polymorphism under controlled temperature and humidity using FT-IR spectroscopic imaging, *Vib. Spectrosc.* 43 (2007) 221e226.
- [38] M.C. Rehbein, S. Husmann, C. Lechner, et al., Fast and calibration free determination of first order reaction kinetics in API synthesis using in situ ATR-FTIR, *Eur. J. Pharm. Biopharm.* 126 (2018) 95e100.
- [39] S. Soldo, A. Trinh, J.D. Kubicki, et al., In situ and real-time ATR-FTIR temperature-dependent adsorption kinetics coupled with DFT calculations of dimethylarsinate and arsenate on hematite nanoparticles, *Langmuir* 36(2020) 4299e4307.
- [40] H. Ren, C. Cai, C. Leng, et al., Nucleation kinetics in mixed NaNO₃/glycerol droplets investigated with the FTIR-ATR technique, *J. Phys. Chem. B* 120(2016) 2913e2920.