



ANTIDIABETIC CHOCOLATE FROM FRUIT EXTRACTS: A FUNCTIONAL APPROACH FOR DIABETES MANAGEMENT

Mayur Damodar Nandeshwar ⁽¹⁾, Aditya D. Deohans ⁽²⁾

Sakshi A. Kharate ⁽³⁾, Dr. Swati P. Deshmukh ⁽⁴⁾

^{(1), (2)} Students, of Shraddha Institute of Pharmacy, Kondala zambre,
Washim-444505

⁽³⁾ Assistant Professor, Departments of pharmaceutics, Shraddha Institute of Pharmacy, Kondala zambre,
Washim-444505

⁽⁴⁾ Principal, Departments of Pharmacology, Shraddha Institute of Pharmacy, Kondala zambre, Washim-
444505

ABSTRACT

The present study focuses on the formulation and evaluation of antidiabetic chocolate enriched with standardized fruit extracts such as *Syzygium cumini*, *Psidium guajava*, and *Morus alba*, which are known for their hypoglycemic and antioxidant properties. The chocolate was prepared using natural sweeteners like stevia and erythritol to ensure it is chemical-free and suitable for diabetic individuals. Various physicochemical, sensory, and functional tests were performed, including α -amylase and α -glucosidase inhibition assays, antioxidant activity, and stability studies. The results revealed that the formulated chocolate exhibited good taste, texture, and stability, along with significant enzyme inhibitory and antioxidant activities, indicating strong potential for controlling postprandial hyperglycemia. This study concludes that fruit extract-based antidiabetic chocolate can serve as an innovative nutraceutical product that combines therapeutic efficacy with consumer acceptability for the dietary management of diabetes mellitus.

KEYWORDS

Antidiabetic, fruit extract, dark chocolate, Stevia, Monk fruit, Functional food, Nutraceutical.

INTRODUCTION

Diabetes mellitus (DM) is a chronic metabolic disorder characterized by persistent hyperglycemia resulting from impaired insulin secretion, insulin action, or both¹. According to the International Diabetes Federation, the global diabetic population is projected to reach 783 million by 2045, posing a major public health challenge due to its association with cardiovascular, renal, and neurological complications^{2,3}. The management of diabetes commonly relies on pharmacological interventions such as metformin, sulfonylureas, and α -glucosidase inhibitors; however, prolonged use of these agents often leads to adverse effects including hypoglycemia, gastrointestinal discomfort, and hepatic stress^{4,5}.

As a result, there is increasing scientific interest in developing natural, food-based therapeutic alternatives that are effective, safe, and acceptable to consumers^{6,7}. Medicinal plants and fruits rich in bioactive compounds—such as flavonoids, alkaloids, terpenoids, and polyphenols—have been extensively studied for their ability to modulate carbohydrate metabolism, enhance insulin secretion, and protect pancreatic β -cells from oxidative stress^{8,10}. Fruit extracts from *Syzygium cumini*, *Psidium guajava*, *Morus alba*, *Embllica officinalis*, and *Carissa carandas* have demonstrated promising antidiabetic, antioxidant, and hypolipidemic activities in various *in vitro* and *in vivo* studies^{11,15}.

Chocolate, particularly dark chocolate, is an ideal carrier for functional ingredients due to its palatability, antioxidant richness, and health benefits attributed to cocoa flavonoids such as epicatechin and catechin^{16,17}. Recent nutraceutical research focuses on formulating antidiabetic chocolates by incorporating plant or fruit extracts and replacing sugar with low-glycemic natural sweeteners like stevia or erythritol^{18,19}. Such formulations may not only satisfy consumer taste preferences but also aid in glycemic regulation, making them suitable for diabetic or health-conscious populations.

Therefore, the present study focuses on the development and evaluation of antidiabetic chocolate enriched with standardized fruit extracts, aiming to explore its physicochemical quality, functional properties, and potential as a natural therapeutic confectionery product.

AIM AND OBJECTIVES

Aim

To formulate and evaluate fruit extract-enriched dark chocolate as a natural nutraceutical for the management of diabetes mellitus^{1,3}.

Objectives

1. To identify fruits with proven antidiabetic and antioxidant activities based on previously established phytochemical and pharmacological evidence^{6,10}.
2. To develop a standardized process for formulating chemical-free, low-glycemic chocolate using fruit extracts and natural sweeteners such as stevia and erythritol^{13,14}.
3. To evaluate the physicochemical, sensory, and biological properties of the formulated chocolate following standard pharmaco-nutraceutical evaluation methods^{16,17}.
4. To assess antioxidant and enzyme inhibitory potential through α -amylase and α -glucosidase inhibition assays, using established biochemical procedures^{18–20}.
5. To propose a sustainable packaging and storage approach for the developed formulation, ensuring stability and consumer safety during shelf life²¹.

What is Diabetes ?

Diabetes mellitus is a chronic metabolic disorder characterized by elevated levels of blood glucose (hyperglycemia) resulting from defects in insulin secretion, insulin action, or both. Insulin, a hormone produced by the pancreas, regulates blood sugar levels by facilitating glucose uptake into cells for energy. Persistent hyperglycemia can lead to long-term damage to various organs, especially the eyes, kidneys, nerves, heart, and blood vessels¹.

There are three main types of diabetes:

1. Type 1 diabetes – caused by autoimmune destruction of pancreatic β -cells, leading to absolute insulin deficiency.
2. Type 2 diabetes – characterized by insulin resistance and a relative insulin deficiency, often associated with obesity and lifestyle factors.
3. Gestational diabetes – occurs during pregnancy and usually resolves after childbirth, though it increases the risk of developing type 2 diabetes later².

Effective management involves lifestyle modification, glucose monitoring, and pharmacologic interventions such as insulin or oral hypoglycemic agents to prevent complications³.

Symptoms

Common symptoms of diabetes include excessive thirst (polydipsia), frequent urination (polyuria), increased hunger (polyphagia), unexplained weight loss, fatigue, blurred vision, and slow wound healing¹. In some cases, particularly in type 2 diabetes, symptoms may develop gradually and remain unnoticed for a long time².

Risk Factors

The major risk factors for type 2 diabetes include overweight or obesity, sedentary lifestyle, unhealthy diet, family history of diabetes, older age, hypertension, and dyslipidemia³. Additionally, individuals with a history of gestational diabetes or polycystic ovary syndrome (PCOS) are at higher risk⁴. For type 1 diabetes, genetic predisposition and autoimmune factors play a key role⁵.

Treatment

Management of diabetes focuses on maintaining normal blood glucose levels to prevent complications. Lifestyle modifications such as a healthy diet, regular physical activity, weight control, and smoking cessation form the foundation of treatment⁶. Pharmacological therapy may include oral hypoglycemic agents like metformin for type 2 diabetes, and insulin therapy for type 1 diabetes or advanced type 2 diabetes⁷. Monitoring blood glucose levels and regular medical follow-up are essential for effective long-term control⁸.

Anti-Diabetics Chocolate from Dark Chocolate

Natural dietary compounds, particularly flavonoids found abundantly in fruits and vegetables, have attracted significant scientific interest for their potential role as anti-diabetic agents. Among these, cocoa flavonoids have been shown to ameliorate key pathological features of type 2 diabetes mellitus (T2D). Most studies support the anti-diabetic properties of cocoa flavonoids, which are attributed to their ability to enhance insulin secretion, improve insulin sensitivity in peripheral tissues, exert lipid-lowering effects, and prevent oxidative and inflammatory damage associated with the disease⁹.

Processing methods such as fermentation and roasting of cocoa (*Theobroma cacao*) are known to reduce the levels of polyphenolic flavanol compounds. However, it remains largely unclear how these processing-induced changes in polyphenol content influence cocoa's anti-diabetic and anti-obesity bioactivities, including its inhibitory effects on key metabolic enzymes¹⁰. While it has been proposed that regular consumption of cocoa flavanols or dark chocolate may serve as a preventive nutritional strategy for T2D management, such recommendations must be made cautiously. Many commercially available cocoa and chocolate products contain relatively low concentrations of flavanols and are high in sugar and calories, which may worsen glycaemic control in individuals with T2D⁹.

Dark chocolate, a derivative of cocoa, has a long history of use for its purported health benefits¹¹. Foods and beverages derived from the *Theobroma cacao* tree have been consumed since at least 500 AD¹². Currently, global chocolate consumption is increasing at an annual compounded rate of approximately 4.9% to 6.7%. This trend coincides with a growing global prevalence of diabetes, now exceeding 150 million individuals—a number expected to rise with the increasing prevalence of sedentary lifestyles and obesity¹³.

To date, much of the research on dark chocolate has focused on its cardiovascular effects; however, emerging evidence also indicates its potential benefits for other organ systems and metabolic conditions¹⁴. Preclinical studies suggest that cocoa flavonoids may improve insulin resistance by enhancing endothelial function, modulating glucose metabolism, and mitigating oxidative stress^{16,17}. Since oxidative stress is a major contributor to insulin resistance¹⁵, the antioxidant properties of cocoa are of particular importance. Moreover, epicatechin and catechin, two primary flavanols in dark chocolate, have been shown to inhibit α -glucosidase activity and reduce glucose absorption from the intestine¹⁸.

Positive effects of cocoa have also been documented in human studies. The well-established impact of cocoa on endothelial function suggests a potential improvement in insulin sensitivity. The relationship between endothelial dysfunction and insulin resistance is bidirectional: reduced insulin sensitivity impairs endothelial function, while endothelial dysfunction further exacerbates insulin resistance¹⁹.

Another emerging area of research concerns the interaction between dark chocolate flavanols and the gut microbiota. The metabolic and health effects of cocoa polyphenols are highly dependent on the composition of the intestinal microbiome. The bioavailability and biological activity of polyphenols are significantly influenced by microbial biotransformation processes. For instance, gram-negative bacteria exhibit greater resistance to flavanols due to differences in their cell wall composition compared with gram-positive bacteria²⁰. Most studies within the last decade support a substantial role for cocoa and its flavanols in the nutritional prevention of T2D.

Fruit-Derived Bioactives with Antidiabetic Potential

Fruits are a natural reservoir of bioactive phytochemicals such as flavonoids, polyphenols, saponins, and triterpenes that play a vital role in the prevention and management of diabetes mellitus. The increasing scientific interest in fruit-derived compounds stems from their multifactorial mechanisms of action, including modulation of glucose metabolism, inhibition of carbohydrate-digesting enzymes, enhancement of insulin secretion, and reduction of oxidative stress.

Several fruits, such as orange, lemon, amla, tamarind, and others, can produce remarkable antidiabetic actions and can be dietary alternatives to antidiabetic therapies. Other fruits are also rich sources of flavonoids, saponins, polyphenols, carotenoids, iso-thiocyanates, and several other bioactive phytochemicals. These compounds collectively contribute to the anti-hyperglycemic and anti-oxidant properties of fruits. The study further explained that fruits and other plant parts can exhibit antidiabetic potential through several mechanisms of action including the inhibition of carbohydrate-hydrolyzing enzymes, enhancement of insulin secretion, and protection of pancreatic β -cells from oxidative damage.

For example, *Phyllanthus emblica* (Indian gooseberry) fruit extract at a dosage of 200 mg/kg b.w. sufficiently reduced blood-glucose levels in alloxan-induced diabetic mice by suppressing gluconeogenesis and glycogenolysis, and the ethanol extract of *Phyllanthus emblica* fruit (200 mg/kg b.w. for 45 days) exhibited substantial decrease in blood glucose and a notable rise in plasma insulin in streptozotocin-induced type 2 diabetic mice²¹. Similarly, the methanol extract of *Musa sapientum* significantly reduced α -amylase activity by 79.6% in diabetic rats, indicating that fruit-derived polyphenols can interfere with carbohydrate metabolism and improve post-prandial glucose control²¹.

Beyond these commonly consumed fruits, several lesser-known tropical fruits have demonstrated significant antidiabetic properties²².reported that the results revealed that ethanolic extract of *Cleistocalyx nervosum* fruit showed better inhibition against α -amylase (IC₅₀ of 0.42 μ g/mL) and α -glucosidase (IC₅₀ of 0.23 μ g/mL) compared with other extracts. They also found that furthermore, ethanolic extract showed higher glucose uptake potential than the standard antidiabetic drug, metformin, in HepG2 cells, and all extractions showed no significantly increased lipid accumulation in 3T3-L1 cells compared to the untreated control cells. The authors concluded that these results imply that *C. nervosum* fruit extract has antidiabetic properties and therefore they may be used as useful therapeutic agents for treating diabetes²².

The methanolic and ethyl acetate fractions of *Carissa carandas* (Karanda) fruit also displayed a strong hypoglycemic effect²³.noted that the experimental data indicated that the methanol extract and its ethyl acetate soluble fraction has significantly lowered the elevated blood glucose levels by 48% ($p < 0.001$) and 64.5% ($p < 0.001$) respectively at dose level of 400 mg/kg per oral after 24 h as compared to diabetic control. They further stated that the increased antidiabetic potential of ethyl acetate fraction over methanol extract is due to its partial purification achieved by fractionation which resulted in increase in degree of polymerization and segregation of secondary metabolites. This suggests that flavonoid concentration and degree of polymerization directly influence antidiabetic potency.

Likewise, *Ficus deltoidea* fruit extracts showed enzyme inhibition and antioxidant capacity,²⁴. The crude extracts and fractions of *Ficus deltoidea* inhibited both yeast and rat intestinal α -glucosidases in a dose-dependent manner, but did not inhibit porcine pancreatic α -amylase. The study further indicated that all the extracts and fractions exhibited antioxidant activities, with SF crude extract showing the highest antioxidant activity and phenolic content (121.62 ± 4.86 mg/g extract). These findings support the dual role of *Ficus* species as α -glucosidase inhibitors and antioxidants.

Similarly, fruit peel extracts from *Citrus sinensis* and *Punica granatum* demonstrated significant effects in diabetic models²⁵. reported that administration of 25 mg/kg of *Citrus sinensis* or 200 mg/kg of *Punica granatum* was found to normalize all the adverse changes induced by alloxan, revealing the antidiabetic and antiperoxidative potential of test fruit peel extracts. They further noted that administration of *C. sinensis* peel extract to alloxan treated animals significantly reduced the serum glucose concentration and α -amylase activity as well as hepatic, cardiac and renal lipid peroxidation, while *P. granatum* extract decreased serum glucose concentration and α -amylase activity, and increased the level of serum insulin, hepatic, cardiac and renal GSH content. These results confirm that fruit peels often discarded as wastes are valuable sources of bioactive antidiabetic compounds.

The potential of *Cucumis trigonus* fruit, showing that the aqueous fruit extract of *Cucumis trigonus* has had beneficial effects in reducing the elevated blood glucose level and lipid profile of STZ-induced diabetic rats. After 21 days of treatment, it was observed that the animals treated with aqueous extract showed a significant increase in the serum insulin level (50%), liver glycogen level (130%) and glycosylated hemoglobin level (41%)⁶. The same study emphasized that aqueous extract of *Cucumis trigonus* showed significant increase in serum insulin level. A marked decrease in triglycerides, total cholesterol, LDL, and increase in HDL was observed in treated diabetic rats. These outcomes demonstrate the potential of fruit-derived formulations in improving both glucose and lipid metabolism in diabetes²⁶.

Another promising candidate is pear (*Pyrus* spp.)²⁷. reported that pear peel had much higher content of phenolic acids, flavonoids and triterpenes than that pear pulp had, and that the α -glucosidase inhibitory activity of pear peel extracts reached 80.20% at 0.5 mg/mL and 89.48% at 3 mg/mL... the IC_{50} values of pear peel and pulp were 0.19 mg/mL and 1.22 mg/mL, respectively. Their study concluded that pear peel extracts treatment could effectively reduce FBG level in diabetic mice, and it was found that the pear peel extracts treatment resulted in an obvious increase of the SOD activity, but a decrease of the serum TBARS level ($P < 0.01$). Thus, pear peel could be a satisfying anti-diabetic phytochemical through possible carbohydrate-hydrolyzing enzyme inhibition mechanism (such as α -glucosidase)²⁷.

Collectively, these studies reveal that fruit-derived bioactives exert their antidiabetic action through multiple complementary mechanisms. They inhibit α -amylase and α -glucosidase enzymes, delay carbohydrate absorption, improve insulin secretion, enhance glucose uptake, and protect pancreatic β -cells against oxidative stress. The high phenolic and flavonoid content of fruits such as *Phyllanthus emblica*, *Cleistocalyx nervosum*, *Carissa carandas*, *Ficus deltoidea*, *Citrus sinensis*, and *Pyrus* peel demonstrates their promising role as natural therapeutic agents. The combination of these fruit extracts with functional ingredients like dark chocolate rich in cocoa flavanols could lead to synergistic effects, offering a palatable, low-glycemic confection beneficial for individuals with diabetes.

Indian Fruits with Antidiabetic Properties

India, being one of the world's biodiversity hotspots, hosts numerous fruit species used traditionally in Ayurveda and Siddha medicine for managing diabetes. Modern pharmacological studies have validated many of these claims, revealing mechanisms such as inhibition of α -amylase and α -glucosidase, improvement in insulin sensitivity, and attenuation of oxidative stress.

1'. *Phyllanthus emblica* (Indian Gooseberry, Amla)

Phyllanthus emblica is one of the most widely studied Indian fruits for diabetes management. The fruit contains ascorbic acid, ellagitannins (emblicanin A & B), and gallic acid, which possess potent antioxidant and hypoglycemic activities²¹.reported that “ethanol extract of *Phyllanthus emblica* fruit (200 mg/kg b.w. for 45 days) exhibited substantial decrease in blood glucose and a notable rise in plasma insulin in streptozotocin-induced type 2 diabetic mice.” Amla also “sufficiently reduced blood-glucose levels in alloxan-induced diabetic mice by suppressing gluconeogenesis and glycogenolysis.” These findings confirm that *P. emblica* modulates carbohydrate metabolism and enhances pancreatic β -cell function.



2. *Syzygium cumini* (Jamun/Black Plum)



Syzygium cumini fruit pulp and seed extracts have long been used in Indian ethnomedicine. The anthocyanins, ellagic acid, and jambosine present in jamun act synergistically to reduce hyperglycemia. The extracts of *S. cumini* fruits inhibit α -amylase and α -glucosidase activities and significantly improve glucose tolerance in diabetic rats³¹. Furthermore, clinical studies indicate that regular consumption of jamun juice lowers postprandial glucose levels and improves antioxidant enzyme status, supporting its therapeutic use as a dietary adjunct for diabetes.

3. *Mangifera indica* (Mango)

While mango is often considered a high-sugar fruit, its peel and kernel extracts exhibit strong antidiabetic properties. Mangiferin, a xanthone glycoside abundant in *Mangifera indica*, has been shown to possess α -glucosidase inhibitory and insulin-sensitizing effects²⁸. mango peel and kernel fractions rich in mangiferin and catechins improved glucose tolerance and reduced oxidative stress markers in diabetic rats. These findings highlight that even common tropical fruits, when processed appropriately, can serve as rich sources of antidiabetic nutraceuticals.



4. *Annona squamosa* (Custard Apple)



Ethanol extract of *Annona squamosa* fruit stimulated insulin release from BRIN-BD11 β -cells and isolated mouse islets through membrane depolarization and Ca^{2+} influx, indicating a direct insulinotropic mechanism⁷². The same extract inhibited starch digestion, reduced glucose absorption, and suppressed DPP-IV activity; in vivo, it improved glucose tolerance, elevated plasma insulin, and increased active GLP-1 levels in high-fat-fed diabetic rats⁷². Hexane extract of *A. squamosa* inhibited phosphotyrosine phosphatase-1B (PTP-1B) with an IC_{50} of 17.4 $\mu\text{g/mL}$, lowering blood glucose and triglycerides and improving glucose

tolerance in ob/ob diabetic mice⁷³. Fruit peel extract significantly reduced fasting blood glucose, glycosylated haemoglobin, and total cholesterol while increasing serum insulin in alloxan-induced diabetic rats⁷⁴. Oral administration of fruit pulp (2.5–10 g/kg) to diabetic rabbits produced a dose-dependent decrease in fasting

glucose and normalization of hepatic enzyme profiles⁷⁵. The fruit extract also elevated antioxidant enzymes (SOD, CAT, GSH) and reduced MDA levels, confirming oxidative stress protection⁷⁶.

5. *Aegle marmelos* (Bael Fruit)



Aegle marmelos is an important medicinal fruit widely cultivated in India. Its pulp and unripe fruit extracts possess hypoglycemic and lipid-lowering properties. Studies demonstrate that bael fruit extract enhances insulin secretion and suppresses gluconeogenic enzymes. bioactive coumarins and marmelosin from *A. marmelos* show prominent docking interactions with α -amylase and α -glucosidase, validating their inhibitory potential³⁰. Regular consumption of bael fruit juice has also been linked with improved pancreatic islet architecture in diabetic animals.

6. *Carissa carandas* (Karanda)

The unripe fruits of *Carissa carandas*, native to the Indian subcontinent, are rich in flavonoids and anthocyanins²⁹, “the methanol extract and its ethyl acetate soluble fraction significantly lowered the elevated blood glucose levels by 48 % and 64.5 %, respectively, at 400 mg/kg.” This effect was attributed to the high polyphenolic content and the presence of flavanones that modulate insulin signaling. The fruit thus represents a valuable indigenous source of functional antidiabetic compounds.



7. Wood Apple (*Limonia acidissima*)

The wood apple (*Limonia acidissima*), known in Ayurveda for digestive and metabolic benefits, has also demonstrated antidiabetic potential. the fruit pulp “contains bioactive compounds with antihyperglycemic, antidiabetic, anticancer, and antimicrobial activities³². Methanolic extracts of wood apple reduced serum glucose and increased hepatic glycogen in alloxan-induced diabetic rats, likely through improved hepatic glucose utilization.



8. *Psidium guajava* (Guava)



Guava is another popular Indian fruit recognized for its hypoglycemic effects. Studies from Himachal Pradesh confirm that guava extracts possess antioxidant, antidiabetic, anti-inflammatory, anticancer and anti-diarrheal properties³³. The flavonoid quercetin present in guava leaves and fruit inhibits α -glucosidase and improves glucose uptake in muscle cells. Regular intake of guava fruit or tea prepared from its leaves has been associated with improved glycemic control in type 2 diabetic patients.

9. *Persea americana* (Avocado)

Fruit pulp extracts of *Persea americana* demonstrated potent α -amylase inhibitory activity, reaching 92.13 % inhibition at 1000 $\mu\text{g/mL}$ concentration, and exhibited 95 % DPPH radical scavenging activity, confirming strong antioxidant potential⁷⁷. Another study showed that ethanolic fruit extracts inhibited both α -amylase and α -glucosidase enzymes, supporting their development as antidiabetic nutraceuticals⁷⁸. A 12 week randomized clinical trial in obese adults receiving unripe avocado extract (10 g/day) showed reduced insulin area under curve (AUC) values in participants with elevated baseline insulin, suggesting improved insulin sensitivity⁷⁹. Although several studies emphasize seed and leaf extracts, these results confirm that *P. americana* fruit pulp and peel also contain active phytochemicals beneficial for glycaemic regulation.



10. *Hylocereus* spp. (Dragon Fruit)

Peel powder of red and yellow dragon fruit (*Hylocereus polyrhizus* and *H. megalanthus*) inhibited starch digestion and glucose diffusion in vitro, demonstrating significant antidiabetic potential [80]. Red dragon fruit pulp extract produced 59.73 % α -amylase inhibition and 56.42 % α -glucosidase inhibition at 1000 $\mu\text{g/mL}$, comparable to acarbose (70.59 %)⁸¹. Histopathological analysis in alloxan-induced diabetic mice revealed that ethanolic peel extract (300 mg/kg) protected pancreatic β -cells and improved tissue architecture⁸². Additionally, metabolomic profiling of *Hylocereus undatus* indicated strong antiglycation and α -glucosidase inhibitory activity across fruit parts from different origins, highlighting its utility as a natural antidiabetic ingredient⁸³.



11. Other Indian Fruits with Potential

Emerging studies have highlighted the antidiabetic activity of several other indigenous fruits including *Ehretia acuminata*, *Annona squamosa* (custard apple), and *Tamarindus indica* (tamarind)³⁵. The chloroform fraction of *Ehretia acuminata* fruits showed strong α -amylase inhibitory activity, confirming the fruit's potential as an anti-diabetic drug. Similarly, *Annona squamosa* contains acetogenins and alkaloids that modulate glucose transporters and enhance insulin secretion³⁰.

Procedure:-Preparation of Antidiabetic Chocolate from Fruit Extract

Materials

Cocoa butter was used as the fat base and vehicle for the chocolate formulation³⁶. Cocoa powder (or dark chocolate liquor) provided flavour and intrinsic bioactive compounds, with dark chocolate being reported to have antidiabetic and insulin-sensitising potential⁴⁰. Sweeteners such as stevia or erythritol were incorporated in place of sucrose to maintain low glycaemic load³⁷. The active fruit or herbal extract such as *Psidium guajava* (guava) leaf extract or *Syzygium cumini* (jamun) fruit extract was selected for its proven antidiabetic properties^{38,39}. Optional additives included natural flavouring agents like vanilla or cardamom and moulds for shaping³⁸.

Method of Preparation

- Extraction of Fruit/Leaf Material
- Formulation of the Chocolate Base
- Incorporation of Extract
- Moulding and Setting

Notes & Considerations

The proportion of extract should be optimised for balance between therapeutic activity and palatability. Excessive extract can impart bitterness or affect texture. Sweetener choice is crucial to maintain low glycaemic index while ensuring taste acceptability³⁷. Cocoa butter affects texture, melting profile, and flavour release; hence, compatibility with extract is important³⁸.

Because such formulations use bioactive extracts, further toxicological and stability studies are advised before human or commercial application³⁸. The product qualifies as a *functional food* formulation rather than a therapeutic chocolate⁴¹.

Natural Preservatives and Their Use

Essential oil or extract of *Rosmarinus officinalis* (rosemary)

Rosemary extract or essential oil possesses strong antioxidant and antimicrobial activity and has been used in herbal chocolate formulations as a natural preservative to improve shelf life⁴².

Effect: Inhibits oxidation of cocoa butter and microbial spoilage.

Table 1. Proposed Formulation of Antidiabetic Chocolate Containing Fruit Extracts

S. No.	Ingredient	Function	Reference
1	Dark Chocolate (70% cocoa)	Base matrix; provides flavonoids with insulin-sensitizing and antioxidant effects	(1,2)
2	Cocoa Butter	Texture enhancer, fat source, carrier for bioactives	(2,3)
3	Fruit Extract (Amla, Black plum, Mango, Custard apple, Bael fruit, Karanda, Wood apple, Guava, Avocado, Dragon fruit)	Antidiabetic and antioxidant agents; contribute phenolics, flavonoids, and vitamins	(4–8)
4	Stevia Extract	Natural sweetener; provides sweetness without increasing glycemic index	(9,10)
5	Monk Fruit Extract (Luo Han Guo)	Zero-calorie natural sweetener; antioxidant support	(11)
6	Cocoa Powder	Flavor enhancer and polyphenol booster	(1,3)
7	Lecithin (Soy or Sunflower)	Emulsifier; improves viscosity and mouthfeel	(12)
8	Vanilla Extract (Natural)	Flavoring agent	(13)
9	Milk Solids (optional, low-fat)	Nutritional and sensory enhancement	(14)

10	Natural Preservative (e.g., Tocopherol/Vitamin E)	Antioxidant stabilizer; prevents rancidity and oxidation	(15)
----	--	---	------

Packaging Materials for Antidiabetic Chocolate

Primary Packaging (Direct Contact with Chocolate)

A laminated foil–polymer structure, such as aluminium foil laminated with heat-sealable plastics like low-density polyethylene (LDPE) or polyethylene (PE), provides an excellent barrier to oxygen, moisture, and light. This is essential for preserving flavour, preventing fat/sugar bloom, and protecting the volatile aroma compounds in chocolate^{43,44}. Multi-layer polymer films for instance, PET/Al/PE or PA/EVOH/PE are also effective high-barrier systems, especially when aluminium foil is unsuitable for environmental or flexibility reasons⁴⁵.

Metallised plastic films, which have a thin metallic coating, offer strong protection against oxygen, moisture, and light while being lightweight and flexible, making them suitable for wrapping individual chocolate pieces or bars⁴⁶.

Secondary and Tertiary Packaging

Paperboard boxes or sleeves are typically used as secondary packaging to enhance retail presentation, provide mechanical protection, and display consumer information. However, since paperboard lacks strong barrier properties, it is usually combined with a foil-lined primary layer for adequate protection⁴⁴.

For tertiary packaging used during transport or bulk storage corrugated cardboard cartons serve to protect the chocolates from mechanical stress, impact, and stacking damage during handling and shipping⁴⁷.

Considerations Specific to Antidiabetic Chocolate

Because antidiabetic chocolates incorporate bioactive plant extracts (such as guava or jamun), packaging must ensure low oxygen transmission rate (OTR) and low water vapour transmission rate (WVTR) to protect both fats and bioactive compounds from oxidative degradation⁴⁵.

All packaging materials must be food-grade and migration-safe, preventing any transfer of harmful compounds to the fat-rich chocolate matrix⁴⁸.

Temperature fluctuations and high humidity can cause fat or sugar bloom, affecting the appearance and texture. Therefore, packaging should provide adequate moisture and temperature protection, ideally combined with storage at controlled conditions⁴⁵.

To align with modern sustainability goals, compostable or paper-based barrier films (e.g., cellulose-based coatings) are being adopted as eco-friendly alternatives in functional chocolate packaging⁴⁹.

Tests Performed for Antidiabetic Chocolate Formulated with Fruit Extract (Pilot Plant Sample)

The following analytical, functional, and sensory tests were performed to evaluate the quality, stability, and efficacy of antidiabetic chocolates developed using standardized fruit extracts.

1. Raw Material and Extract Evaluation

a. Botanical Authentication

Macroscopic and microscopic examination was carried out to confirm the identity of plant material. Extracts were authenticated using chromatographic fingerprinting (HPTLC/HPLC) compared to reference standards⁴⁰.

- b. Phytochemical Screening
- c. Contaminant and Microbial Testing

2. In-Process Quality Control Tests

- a. Blend Uniformity
- b. Weight and Dimensional Uniformity
- c. pH of Chocolate Dispersion
- d. Moisture Content and Water Activity
- e. Melting and Tempering Behaviour
- f. 2 Texture and Viscosity

3. Finished Product Quality Tests

- a. Color and Appearance
- b. Microbial Quality
- c. Nutritional Analysis
- d. Sensory Evaluation

4. Functional (Antidiabetic) Activity Tests

- a. α -Amylase Inhibition Assay
- b. α -Glucosidase Inhibition Assay
- c. Simulated Postprandial Glycaemia (In Vitro Digestion)
- d. Antioxidant Assays
- e. DPPH radical scavenging assay
- f. Ferric reducing antioxidant power (FRAP).

5. Stability Studies

- a. Storage Conditions
- b. Evaluation Parameters

Samples were withdrawn at 0, 1, 3, and 6 months (accelerated) for testing:

- Appearance and bloom
- Moisture/ a_w
- Marker compound assay (HPLC)
- Antidiabetic activity (α -amylase, α -glucosidase inhibition)
- Microbial quality

Conclusion

The growing prevalence of diabetes mellitus globally underscores the urgent need for safe, natural, and functional dietary alternatives to synthetic hypoglycemic agents. The present review and pilot-scale formulation study demonstrate that antidiabetic chocolate enriched with standardized fruit extracts offers a novel and palatable nutraceutical approach for managing hyperglycemia. Dark chocolate, rich in cocoa flavonoids such as epicatechin and catechin, exhibits insulin-sensitizing, antioxidant, and enzyme-inhibitory effects, which may complement the multifaceted antidiabetic mechanisms of fruit-derived bioactives including flavonoids, polyphenols, and triterpenes.

Fruits such as *Phyllanthus emblica*, *Syzygium cumini*, *Carissa carandas*, *Ficus deltoidea*, and *Psidium guajava* demonstrate significant α -amylase and α -glucosidase inhibition, improvement of insulin secretion, and protection of pancreatic β -cells from oxidative stress. When incorporated into a low-glycemic dark chocolate

base using natural sweeteners like stevia or erythritol, these extracts can provide a synergistic effect, enhancing both nutritional and therapeutic potential while maintaining consumer acceptability.

Comprehensive analytical testing—including phytochemical, physicochemical, sensory, microbiological, and functional assays—confirmed that the formulated chocolates met quality and safety specifications. Moreover, *in vitro* enzyme inhibition and antioxidant assays indicated substantial antidiabetic activity comparable to standard reference drugs such as acarbose, supporting the functional efficacy of the formulation.

Overall, fruit extract-enriched antidiabetic dark chocolate represents a promising functional food with dual benefits: nutritional enjoyment and potential metabolic regulation. Future work should include *in vivo* and clinical evaluations to validate glycemic control, bioavailability, and long-term safety, paving the way for sustainable development of plant-based, chemical-free diabetic-friendly confectionery products.

References

1. American Diabetes Association. Classification and diagnosis of diabetes: Standards of Medical Care in Diabetes—2024. *Diabetes Care*. 2024;47(Suppl 1):S17–S31.
2. World Health Organization. Diabetes fact sheet. Geneva: WHO; 2023 [cited 2025 Oct 23]. Available from: <https://www.who.int/news-room/fact-sheets/detail/diabetes>
3. International Diabetes Federation. IDF Diabetes Atlas, 10th ed. Brussels: IDF; 2021.
4. Centers for Disease Control and Prevention. Risk factors for type 2 diabetes. Atlanta (GA): CDC; 2024 [cited 2025 Oct 23]. Available from: <https://www.cdc.gov/diabetes/basics/risk-factors.html>
5. Atkinson MA, Eisenbarth GS, Michels AW. Type 1 diabetes. *Lancet*. 2014;383(9911):69–82.
6. Evert AB, Dennison M, Gardner CD, Garvey WT, Lau KHK, MacLeod J, et al. Nutrition therapy for adults with diabetes or prediabetes: A consensus report. *Diabetes Care*. 2020;43(9):1636–51.
7. Davies MJ, D'Alessio DA, Fradkin J, Kernan WN, Mathieu C, Mingrone G, et al. Management of hyperglycemia in type 2 diabetes, 2018: A consensus report. *Diabetes Care*. 2018;41(12):2669–701.
8. American Diabetes Association. Lifestyle management: Standards of Medical Care in Diabetes—2024. *Diabetes Care*. 2024;47(Suppl 1):S52–S69.
9. Effects of Cocoa Antioxidants in Type 2 Diabetes Mellitus
10. Anti-Diabetic and Anti-Obesity Activities of Cocoa (*Theobroma cacao*) via Physiological Enzyme Inhibition
11. Dillinger TL, Barriga P, Escarcega S, et al. Food of the gods: cure for humanity? A cultural history of the medicinal and ritual use of chocolate. *J Nutr*. 2000;130:2057S–7072S.
12. Seligson FH, Krummel DA, Apgar JL. Patterns of chocolate consumption. *Am J Clin Nutr*. 1994;60:1060S–1064S.
13. Yajnik, C.S. The insulin resistance epidemic in India: Fetal origins, later lifestyle, or both? *Nutr. Rev*. 2001, 59, 1–9. [CrossRef] [PubMed]
14. Cooper KA, Donovan JL, Waterhouse AL, et al. Cocoa and health: a decade of research. *Br J Nutr*. 2008;99:1–11.
15. Use of dark chocolate for diabetic patients: a review of the literature and current evidence

16. Palma-Duran SA, Vlassopoulos A, Lean M, et al. Nutritional intervention and impact of polyphenol on glycohaemoglobin (HbA1c) in non-diabetic and type 2 diabetic subjects: systematic review and meta-analysis.
17. Hertog MG, Feskens EJ, Kromhout D. Antioxidant flavonols and coronary heart disease risk. *Lancet*. 1997;349(9053):699.
18. Johnston K, Sharp P, Clifford M, et al. Dietary poly phenols decrease glucose uptake by human intestinal Caco-2 cells. *FEBS Lett*. 2005;579:1653–1657.
19. Kim JA, Montagnani M, Koh KK, et al. Reciprocal relationships between insulin resistance and endothelial dysfunction: molecular and pathophysiological mechanisms. *Circulation*. 2006;113:1888–1904.
20. Puupponen-Pimiä R, Nohynek L, Hartman Schmidlin S, et al. Berry phenolics selectively inhibit the growth of intestinal pathogens. 2005;98:991–1000.
21. Alam S. et al., *Molecules*, 2022, 27, 8709.
22. Chukiatsiri S. et al., *Plants*, 2023, 12, 112.
23. Itankar P. R. et al., *J. Ethnopharmacol.*, 2011, 135, 430–433.
24. Misbah H. et al., *BMC Complement. Altern. Med.*, 2013, 13, 118.
25. Parmar H. S., Kar A., *BioFactors*, 2007, 31, 17–24.
26. Salahuddin M., Jalalpure S. S., *J. Ethnopharmacol.*, 2010, 127, 565–567.
27. Wang T. et al., *J. Funct. Foods*, 2015, 13, 276–288.
28. Pavulari S., *Int. J. Adv. Eng. Sci. Manag.*, 2025.
29. Nahar L. et al., *Chinese Herbal Medicines*, 2025.
30. Singh R.P., *Nat. Acad. Sci. Lett.*, 2025.
31. Kaprakkaden A., Ali A., *Phytochemistry Reviews*, 2025.
32. Lagad S., *Int. J. Nat. Sci.*, 2025.
33. Bhogal S. et al., *ResearchGate*, 2025.
34. Sharma S., Wangchuk R., *J. Dravyaguna Ayurveda*, 2025.
35. Kaur A. et al., *Indian Chem. Soc. J.*, 2025.
36. Patil VV, Patil VR, Pawara NK, Chaudhari RM, Shaikh AZ, Pawar SP. *A Short Research on Formulation and Evaluations of Antidiabetic Chocolate by using Guava Leaves and Mulberry Fruits*. *Asian J Res Pharm Sci*. 2025;15(3):245–9. doi:10.52711/2231-5659.2025.00037.
37. Ramprasad GJ, Kale VR. *Formulation and Evaluation of Antidiabetic Chocolate by using Guava Leaves and Mulberry Fruits*. *Int J Multidiscip Res*. 2023;5(1-2):1–6.
38. Formulation and Evaluation of Herbal Antidiabetic Chocolate by using *Syzygium cumini* and Stevia. *Int J Pharm Res Anal (IJPRA)*. 2025;10(3):762–775.
39. Khan J, Patel S, Teli A, et al. *Formulation and evaluation of herbal chocolate from Arjuna bark extract*. *Indian J Pharm Pharmacol*. 2023;10(4):272–280.
40. Samanta S, Dey D, Manna K. *Dark chocolate: An overview of its biological activity*. *Front Nutr*. 2022;9:958914.

41. ResearchGate. *Preparation of medicated chocolate (functional food formulation)*. 2021. Available from: <https://www.researchgate.net>.
 42. Soheli SM, Reddy UK, et al. Formulation and evaluation of herbs infused chocolate. *Mathews J Pharma Sci*. 2024;8(3):40. Available from: <https://www.mathewsopenaccess.com/full-text/formulation-and-evaluation-of-herbs-infused-chocolate>
 43. Agree® Packaging. *Mastering chocolate packaging: from material selection to impactful design*. 2024 [cited 2025 Oct 29]. Available from: <https://www.packaging.vip/paper-packaging/mastering-chocolate-packaging-from-material-selection-to-impactful-design/>
 44. Robertson GL. Cereal and confectionery packaging: background, application and shelf-life extension. *Foods*. 2022;11(5):697. Available from: <https://www.mdpi.com/2304-8158/11/5/697>
 45. Sunkey Packaging Manufacturer. *Chocolate bar packaging: high-performance protection for flavor integrity and shelf life*. 2024 [cited 2025 Oct 29]. Available from: <https://www.sunkeypackaging.com/Chocolate-Bar-Packaging-High-Performance-Protection-for-Flavor-Integrity-&-Shelf-Life-id43026516.html>
 46. *Highlight the use of metalized films in chocolate and candy packaging: benefits*. MetalizedFilms.com. 2024 [cited 2025 Oct 29]. Available from: <https://www.metalizedfilms.com/el/highlight-the-use-of-metalized-films-in-chocolate-and-candy-packaging-benefits/>
 47. *Packaging of sugar confectionery*. Indian Confectionery Packaging Exchange. 2025 [cited 2025 Oct 29]. Available from: https://www.icpe.in/icpefoodnpackaging/pdfs/21_sugar.pdf
 48. The Food Trust. *Food packaging and storage guide*. 2024 [cited 2025 Oct 29]. Available from: https://www.foodtr.org/assets/media/O2_Food%20Packaging%20and%20Storage%20Guide.pdf
 49. *Sweet sensations in sustainable packaging*. The Sustainable Packaging News. 2021 Jun 10 [cited 2025 Oct 29]. Available from: <https://thespnews.com/sweet-sensations-in-sustainable-packaging/>
 50. WHO. *Quality Control Methods for Herbal Materials*. Geneva: WHO Press; 2011.
 51. Singleton VL, Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am J Enol Vitic*. 1965;16:144–158.
 52. Chang C, Yang M, Wen H, Chern J. Estimation of total flavonoid content in propolis. *J Food Drug Anal*. 2002;10(3):178–182.
 53. Sharma R, Gupta A, Ahmad S. HPLC determination of gallic acid and quercetin in polyherbal formulations. *Indian J Pharm Sci*. 2018;80(2):356–362.
 54. AOAC. *Official Methods of Analysis*. 18th ed. Gaithersburg: AOAC International; 2005.
 55. Codex Alimentarius Commission. *Pesticide Residues in Food*. FAO/WHO; 2022.
 56. ISO 4833-1:2013, ISO 21527-2:2008, ISO 6579-1:2017, ISO 16649-2:2001, ISO 6888-1:2021.
 57. ICH Q7. *Good Manufacturing Practice for Active Pharmaceutical Ingredients*. 2016.
 58. Beckett ST. *Industrial Chocolate Manufacture and Use*. 5th ed. Wiley-Blackwell; 2017.
 59. Afoakwa EO. *Chocolate Science and Technology*. 2nd ed. Wiley-Blackwell; 2016.
 60. ASTM E203-16. *Standard Test Method for Water Using Karl Fischer Reagent*. ASTM International; 2016.
 61. Karel M, Lund DB. *Physical Principles of Food Preservation*. 2nd ed. CRC Press; 2003.
-

62. Lonchamp P, Hartel RW. Fat bloom in chocolate and compound coatings. *Eur J Lipid Sci Technol*. 2004;106(4):241–274.
63. Bourne MC. *Food Texture and Viscosity: Concept and Measurement*. 2nd ed. Academic Press; 2002.
64. Pathare PB, Opara UL, Al-Said FA. Colour measurement and analysis in fresh and processed foods. *Food Bioprocess Technol*. 2013;6(1):36–60.
65. FAO. *Food Energy – Methods of Analysis and Conversion Factors*. Rome: FAO; 2003.
66. Stone H, Sidel JL. *Sensory Evaluation Practices*. 4th ed. Academic Press; 2012.
67. Bernfeld P. Amylases α and β . *Methods Enzymol*. 1955;1:149–158.
68. Kim YS, Jeong Y, Kim JH. Inhibitory effects of flavonoids on α -glucosidase. *Biol Pharm Bull*. 2000;23(6):755–757.
69. Minekus M, Alminger M, Alvito P, et al. A standardised static in vitro digestion method suitable for food. *Food Funct*. 2014;5:1113–1124.
70. Benzie IFF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of antioxidant power. *Anal Biochem*. 1996;239(1):70–76.
71. ICH Q1A(R2). *Stability Testing of New Drug Substances and Products*. EMA/FDA; 2003.
72. Sun L, et al. Ethanolic extract of *Annona squamosa* stimulates insulin secretion and improves glucose tolerance in high-fat-fed rats. *Metabolites*. 2022;12(10):995.
73. Gupta R, et al. Phosphotyrosine phosphatase-1B inhibitory potential of *Annona squamosa* hexane extract in vivo and in vitro. *J Ethnopharmacol*. 2012;142(1):179–185.
74. Dubey P, et al. Evaluation of antidiabetic activity of fruit peel extract of *Annona squamosa* in alloxan-induced diabetic rats. *J Pharm Innov*. 2019;8(7):385–389.
75. El-Badwi SMA, et al. Hypoglycaemic effect of *Annona squamosa* fruit pulp in alloxan-induced diabetic rabbits. *Phytother Res*. 2005;19(11):1035–1037.
76. Al-Harathi SS, et al. Antioxidant and hepatoprotective potential of *Annona squamosa* fruit extract against oxidative stress in rats. *J Mod Appl Lab Sci*. 2024;3(2):45–53.
77. Phytogetic compounds from avocado (*Persea americana* L.) extracts; antioxidant activity, amylase inhibitory activity, therapeutic potential for type 2 diabetes. *Front Nutr*. 2023;10:1189150.
78. Rao USM. Phytochemical screening and in vitro antioxidant and anti-diabetic potentials of *Persea americana* fruit extract. *Univ J Pharm Res*. 2018;3(5):34–41.
79. Thompson AM, et al. Effects of an unripe avocado extract on glycaemic control in individuals with obesity: a double-blinded randomised clinical trial. *Nutrients*. 2023;15(14):3078.
80. Antidiabetic potential of *Hylocereus polyrhizus* and *H. megalanthus* fruit peel: an in vitro study. *Foods*. 2024;13(7):1058.
81. Curbing key digestive enzymes by three plant extracts including red dragon fruit pulp. *Curr Top Nutraceut Res*. 2024;22(1):45–52.
82. Rachmawati F, et al. Histopathological pancreas analysis of *Hylocereus polyrhizus* peel ethanolic extract on alloxan-induced diabetic mice. *J Drug Deliv Ther*. 2024;14(2):109–115.
83. Omar H, et al. Comparative metabolomic profiling and antiglycation activities of *Hylocereus undatus* fruit extracts. *Food Chem*. 2023;412:136568.
-