



Nanopore-Based Theory As Approach In Variables Of Diagnosis

Nanopores ; application and variations

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Abstract: Modern diagnostic tools aim to be precise, quick, and affordable while effectively spotting diseases, infections, or illnesses. Detecting a disease early on is crucial as it can determine the difference between life-threatening outcomes and successful treatment. One difficulty with various diseases is that the ability to detect them increases as the disease advances. Because single molecule sensors like nanopores can detect biomolecules even at low levels, they could be important in medical practices today. Using nanopores for sensing means you need less of a substance to detect it, which can be helpful in clinics. Nanopores can also be adjusted to detect many different types of molecules, are inexpensive, and can be made in large quantities. We talk about past studies and how nanopores can be used in medicine to detect diseases early, track treatments, and recognize when a disease comes back. Infections are a big problem for public health and they are becoming more dangerous for people everywhere. New or familiar recurring germs, or germs that are hard to fight against, make it difficult for us to identify and manage them. Nanopore sequencing technology can help us improve our capability to detect infectious diseases. To diagnose, question, and monitor infectious diseases easily because of the flexible reading length and portable system. This review discusses how nanopore sequencing technology is used in diagnosing infectious diseases in clinical settings. The content covers a quick overview of nanopore sequencing technology and the sequencing platforms by Oxford Nanopore Technologies (ONT). (i) Ways to use nanopore-based sequencing technologies; and (ii) How nanopore sequencing technology is used in keeping track of new developments. Identifying harmful germs, finding specific drug-resistant genes, and studying microbes linked to diseases are important areas of research in microbiology. In this paper, we talk about communities. Then, we look at the challenges, opportunities, and future of communities. The potential use of nanopore sequencing technology in diagnosing infectious diseases is being considered.

Index Terms - Nanopores , sequencing , Mdna , ONT

I. INTRODUCTION

In the 21st century, infectious diseases still create a significant burden on public health care. New harmful microorganisms are always appearing, along with pathogens that are resistant to drugs, and old diseases are coming back in outbreaks. New and difficult challenges have emerged for the technology used to diagnose infectious diseases in recent years. Highly dangerous pathogens with the potential for causing severe illness and spreading quickly have appeared, such as the outbreak of severe acute respiratory syndrome (SARS) in 2002-2003. In 2012, a virus called Middle East respiratory syndrome coronavirus (MERS-CoV) emerged. The Ebola virus was a big concern from 2014 to 2016, and the Zika virus was a problem from 2015 to 2016. The current coronavirus, known as SARS-COV-2, has caused a severe respiratory illness recently. Additionally, pathogens that have developed resistance present significant risks to public health and are a

cause for concern. Many people are talking about it. When we look at how fast antibiotics are being developed, the issue of persistent infections is getting a lot of focus. Drug resistance is rapidly increasing and affecting almost all commonly used medications, even those considered as a last resort, such as colistin and polymyxin B. This has become a serious issue. A new report called the Global issue¹⁴ was just released. The 2019 Burden of Disease Study gave a detailed look at how often diseases occur and how widespread they are. show the number of deaths caused by different diseases and injuries in various countries and territories from 1990 to 2019. New information shows that infectious diseases, such as lower respiratory infections and diarrheal diseases, are the leading reason for disability in children. Malaria, meningitis, whooping cough, and sexually transmitted infections are some common diseases. HIV/AIDS is the second most prevalent. Alternative options are the main reason for disability-adjusted life-years in the age group of 25-49 years. It is very important to have diagnostic tools that can help doctors make more precise and quick diagnoses in medical settings. In the past, old-fashioned ways of testing or tests that look for antigens take a lot of time, need a lot of work, are expensive, and only look at one person at a time. Thankfully, thanks to ongoing improvements in technology, scientists now focus on studying individual pathogens instead of entire populations. In labs and well-equipped clinics, it's pretty common to use molecular biology and detection technologies. New molecular diagnostic technologies like isothermal amplification technology RT-qPCR and MALDI-TOF mass spectrometry are becoming more advanced. Multiplex PCR detection, as well as high-resolution techniques, are utilized for identifying various substances. The ability to diagnose pathogens and drug-resistant genes is quickly changing due to melting technologies. One downside of using these techniques is that they cannot identify unfamiliar pathogens and have high limitations. Despite the positive sensitivity, efficiency, and specificity, expenses remain a challenge. Advances in sequencing technology have been rapid. New opportunities have been made for finding disease-causing germs and drug-resistant genes. NGS technology, when paired with bioinformatics methods, can help find all microorganisms present in a sample. . Regrettably, the sequencing process is limited to 24 hours. Reading DNA sequences that are typically less than 500 base pairs, obtained through advanced technologies like Illumina and Roche 454 sequencing, can be challenging when trying to understand the intricate structure of the genome. Microorganisms, particularly those with numerous repeated elements, are best analyzed using long-read sequencing technology. Technologies like ONT and Pacific Biosciences are examples of advanced tools in the field. Specialists in content writing have successfully dealt with various challenges, resulting in a significant improvement in genome sequencing for clinical diagnosis. ONT has been at the forefront of advancing and bringing to market nanopore-based sequencing technology. When comparing PacBio to nanopore sequencing, nanopore sequencing is a preferable option. The instrument is affordable, easy to carry around, and can read longer sequences. In this article, we give a quick look at how nanopore sequencing technology is used in diagnosing infectious diseases in a clinical setting. Talking about the problems and possibilities of this technology can inspire more research on diagnosing clinical pathogens. different ways to do something Nanopore sequencing platforms work by watching the flow of ions. Changes in levels due to single DNA or RNA molecules moving through a tiny channel or pore. Later on, the changes in ionic current are analyzed using base-calling algorithms to figure out the molecular sequence. In recent years, there has been an increase in the use of nanopore sequencing technology. Various platforms have been created, each having special features suitable for different uses. The MinION, which is small enough to fit in your pocket and runs on USB power, is one such example. ONT released its first commercially-available sequencer in 2014. The MinION nanopore sequencer is portable, so it can be taken out of the lab and used to find pathogens in difficult field conditions. GridION and PromethION sequencers offer greater capacity compared to MinION. This allows for analysis on a bigger scale and a cost-effective method for sequencing large genomes. The first labs to receive PromethION got it in early 2017. It is known for being a very high-capacity platform. After that, the tool was improved, and now the sequencing platform has either 24 or 48. Single flow cells that offer a lot of data (up to 7 or 14 Tb). The affordable Flongle device for fluid flow. The cell sequencing device works as an adapter for MinION and GridION, making it possible to sequence smaller tests and experiments at a lower cost. In the coming years, sequencing will continue to get smaller and be done all at once. New devices are being created that will offer more flexibility. Smaller platforms are currently in the works. ONT also offers SmidgION, which works with smartphones and other mobile devices. The MinION Mk1D is made to be a special keyboard that comes with a built-in sequencer for tablet devices. Furthermore, Plongle is made for people who want to do many fast tests using its 96 disposable flow cells. Parallel technology allows users to achieve a low cost per sample. When compared to traditional short-read technology, parallel technology is more cost-effective. One of the benefits of nanopore sequencing technology is that it provides real-time data streaming, giving quick access to useful information. Results can be seen and the sequencing can be stopped

at any point as long as there is enough sequencing data. The process can be scaled up. The modular MinION, GridION, and PromethION feature is great for sequencing pathogens at various levels, from low to ultra-high throughput. (iii) The ability to easily move and adjust allows... Using portable and affordable MinION devices allows you to get the results you desire, whenever and wherever you need them. You can change both the order of the steps and how many samples are tested. Plus, you have the option to adjust. You can sequence many samples at the same time by using up to 96 barcodes. The length of the reads is also important. Nanopore sequencing is not limited by any restrictions. Technology allows for reading lengthy genetic material, such as complete genomes, plasmids, and long repeat regions, without any restrictions, leading to successful outcomes. (v) The process is made more efficient and automated using Nanopore technology. Tools based on nanopores have simplified and automated processes, cutting down on the time spent on them. This makes nanopore-based tools more convenient for use. Big genetic studies have been done. Also, ONT has come out with a new Kit recently. Chemistry now has a new sequencing enzyme that can achieve accuracies of over 99%. The assembly accuracy of the Oxford Nanopore R10.4 flow cell has greatly improved with the newest Q20+ chemistries. Due to the high error rate of the R9.4.1 flow cell in detecting homopolymers, it is necessary to perform short-read polishing. To improve the assembly outcomes in order to create top-notch genomes, specific adjustments are needed. Barriers have prevented the widespread adoption of nanopore sequencing technology. The newest version from Oxford Nanopore has the potential to overcome these obstacles. The R10.4 flow cell has greatly improved the accuracy of sequencing to around 99%. We can get great microbial genetic information just from R10.4 data when we have 40x coverage from pure cultures or metagenomes, without needing short-read or. . Improving the referencing techniques of Nanopore sequencing. New and powerful methods using nanopore sequencing platforms have been implemented. Whole-genome sequencing is used for various experimental purposes to fulfill different needs. Nanopore sequencing systems can enhance assembly accuracy, which helps researchers study how repeat elements impact microbial function and adaptation. Targeted sequencing is often used to quickly identify microorganisms. In metagenomic analysis, nanopore technology is used to study the mechanisms and detect drug-resistant genes in pathogens. Sequencing technology helps put together full bacterial genomes and plasmids from different metagenomic samples. Examples to offer impartial and PCR-free genetic code sequences. Targeted sequencing is enhanced by the long sequencing reads provided by nanopore technology, which broadens its capabilities. other types of genetic variations, such as indels and structural variants, are becoming increasingly popular. These methods offer more comprehensive and accurate insights into the genetic makeup of individuals. Targeted nanopore sequencing can detect structural variations, repetitive regions, and base modifications in DNA. Focus on collecting data from a specific part of the genetic code to get more precise information about biological traits. Specialists have found agents in complicated samples. It is important to mention. However, this method increases the ability to detect specific microorganisms, it may not cover all possible pathogens that could be present. Nanopore-targeted amplicon sequencing is a method that uses specific primers to amplify certain genes. The technique has limitations. Study of tiny living organisms in medical samples gives us new and important information about the bacteria present. Communities and factors that make drugs less effective. Arranging genes that are related to taxonomy in full, like 16S and 23S for bacteria and 18S for fungi. Enhances the accuracy of identification by using a method that depends on multiplex PCR for specific targeting. By enhancing viral genomes, experts can identify viral consensus sequences within 1-2 days of collecting clinical samples. One more benefit of nanopore targeted sequencing is that it combines targeted amplification and sequencing. Using long-read nanopore sequencing can detect SARS-CoV-2 and other respiratory viruses at the same time in just 6-10 hours. The test could detect as few as 10 standard plasmid copies in each reaction, and it was specifically designed to detect SARS-CoV. The accuracy of Strategy 2 for COVID-19 diagnosis was 100%, making it a suitable method that can be expanded further. Identify other germs. A way to find SARS. Clinical samples are being analyzed for CoV-2 genome sequencing using a method called rapid sequencing long amplicons (RSLAs). This process involves using random primers to create cDNA from RNA that has been purified from the samples. Doctors use clinical samples to conduct single or multiplex PCRs in order to create longer amplicons of the SARS-CoV-2 genome. This method can detect SARS-CoV-2 and offer better accuracy in finding viruses in medical samples that might have been missed as negative. Other methods that use nucleic acids are also used. Sequencing is a technique that combines multiplex amplicon PCR. Using nanopore technology to read drug resistance genes from clinical samples without the need for culturing is a flexible and easy way to diagnose infections. The technique offers high sensitivity and accuracy, giving it an advantage. Additionally, we can find mutations that are not linked to drug resistance and identify new mutations as well. Application has been increasingly used in various fields such as genetics, medicine, and environmental science. Many ways have been studied to

diagnose infectious diseases. The use of Experts have explained high-throughput sequencing thoroughly in various medical areas. Moreover, the increasing use of ONT platforms has led to a shift in focus towards automation. Finding and describing harmful germs quickly and correctly is important for monitoring them. Identifying and treating infectious diseases relies on understanding microorganisms as the outbreak spreads. Right now, the main way to do this is through culture-based methods. Detecting germs in a medical setting is important, but using culture-based methods takes a lot of time and effort, causing delays in diagnosis. In recent years, nucleic acid amplification tests (NAATs) have been used to quickly detect harmful pathogens. Microorganisms can be detected using realtime PCR assays, but these methods have some restrictions. NAAT tests are made using known sequences, so there is a chance of getting results that say something is negative when it's actually positive for organisms with high levels. Detecting unknown or emerging targets using existing assays is not feasible due to mutation rates. Additionally, many NAATs, such as real-time PCR, are unable to capture specific genome sequences. As a result, it may not be possible to conduct a detailed study on how pathogens spread and change over time. Certainly, nanopore sequencing technology addresses these issues. It is a new and improved method. New infectious diseases can be a big problem for public health because they can infect a lot of people. The ongoing COVID-19 pandemic has shown how serious and costly diseases can be. The Ebola outbreak in Guinea was another example of this. In their study, et al. created a sequencing system using the ONT MinION platform. In one regular test setup, realtime monitoring of the epidemic's genome can be done. During the Zika outbreak in Brazil, the team of researchers used PCR and sequencing techniques at Oxford University. A specialist content writer used the Nanopore MinION platform to sequence a group of samples and identify them. Studying and understanding the genetic makeup of ZIKV in infected samples helps in tracing how the virus spread during the epidemic. tracking how it spreads to different areas. Lately, the COVID-19 outbreak caused by SARS-CoV-2 has been a concern. The pandemic had a huge effect all around the world. It is very important to correctly find SARS-CoV-2 for it to be controlled effectively. Controlling and managing the pandemic is crucial. China finished sequencing the genome of the SARS-CoV-2 virus in January 2020 and shared the results with other countries. Changes and mixing of genetic material have happened while SARS-CoV-2 was replicating. Nanopore technology was used to uncover the sequence of the viral genome. Some patients had a certain amount of missing viral genetic material, which might cause incorrect test results when using other genetic tests. acid-based tests are being used. Right now, reverse real-time techniques are being utilized. RT-PCR tests are commonly used in clinics to detect SARS-CoV-2. The genetic material of the virus includes nucleocapsid (N), envelope (E), and genes called open reading frame 1a or 1b. These genes have specific binding sites. If there are changes to the primers and probes in these genes, it will make the assay less accurate and sensitive. Researchers discovered that when using Oxford Nanopore platforms for whole genome sequencing of SARS-CoV-2, they were able to gather valuable information. The nonstructural protein 1 (nsp1) gene is found at the beginning of the SARS-CoV-2 genome and is commonly found in samples taken from the nose or saliva of COVID-19 patients. A new test called nsp1 real-time RT-PCR was included in the detection of SARS-CoV-2 for 19 patients with various levels of illness. This can prevent getting incorrect results because of changes in the parts where the primer or probe attaches in the RT-PCR tests that are used now. Furthermore, getting infected with both SARS-CoV-2 and other respiratory viruses at the same time is also possible. Detecting SARS-CoV-2 is a big challenge because of viruses, as mentioned by Wang and colleagues. Scientists created a new way to detect viruses like SARS-CoV-2 using tiny holes called nanopores. The test can detect multiple viruses within 6-10 hours and can identify as few as 10 standard plasmid copies per test. Nanopore sequencing techniques are useful for identifying pathogens and sequencing genomes, as well as categorizing different types of pathogens. The nanopore sequencing system can detect different kinds of viruses at the same time using the known SARS-CoV-2 GenBank. Researchers can use this technology to follow how diseases spread and how pathogens change over time.

Detection of drug resistance The misuse of drugs has caused a concerning rise in the number of drug resistance genes being detected. Resistance is present in pathogens that lead to infections acquired in healthcare settings and within the community. Diseases caused by drugresistant germs can result in serious illness and even death. This is why it is important to address this issue promptly. There is a requirement for fast and thorough ways to precisely explain particular gene profiles related to drug resistance. Understanding the presence of bacteria-related antimicrobial resistance is seen as necessary in order to treat diseases effectively and promptly. Controlling the spread of AMR is important, but it often takes more than 48 hours for microbiological culture to give results. This delay in diagnosis can cause the problem to spread further. Drug-resistant bacteria are a big problem. Scientists have come up with different ways to detect them using molecular biology techniques. Solving this issue with all AMR causes and finding new mutations is not possible using the current method. Nanopore analysis is a growing field that can help with

this. Many different fields have a strong interest in detecting genes that cause antibiotic resistance. Gonorrhea is the second most frequently occurring sexually transmitted infection. Bacterial infections are spread all over the world and are resistant to antibiotics. The bacterium *Neisseria gonorrhoeae*, also known as NG, causes gonorrhea and makes it hard to stop and control this disease. It is a big concern for global health. worry 88,89 Our team successfully acquired the entire genetic code of *N. gonorrhoeae* using the MinION sequencer, marking the first time this has been achieved. In contrast, many of the AMR characteristics found with nanopore-based assembly were similar to those of the AMR. Hybrid assemblies have found determinants by combining Illumina and MinION reads in a specific order. You can find AMR profiles for seven different types of antibiotics by using Pathogenwatch and following a BLASTbased process. In addition, a new change related to fighting germs was discovered in *mtrR*, suggesting that this test could be useful. One way to find new AMR causes is through a special method called multiplex PCR amplicon nanopore. Using a bioinformatics analysis process along with sequencing helps to better find AMR genes in clinical samples. Using this plan, our team developed a new technique that I was able to sequence 13 genes related to antibiotic resistance in *N. gonorrhoeae* all at once, starting from clinical samples. Samples can be analyzed within a time frame of 7 hours and 40 minutes to 10 hours and 40 minutes. This is faster than Sanger sequencing. In tests, this method showed an accuracy of more than 99.5% and correctly identified the AMR sites. Additionally, this approach allows for finding AMR genes in pathogens that are not well studied, like *Mycobacterium*. In the 2020 Global Tuberculosis Report released by the World Health Organization, they discussed tuberculosis (MTB). According to the World Health Organization (WHO), around 10 million people got sick with tuberculosis (TB) in 2019. Drug-resistant MTB, which causes TB, is difficult to treat. Many patients with TB face this challenge. In one research, TB was found to be resistant to multiple drugs. We improved the TB testing kit and drug resistance testing tool called Deeplex Myc-TB for better results. The researchers tested whether the product designed for Illumina sequencing could also be used on the ONT MinION sequencer. Research showed that Deeplex Myc-TB can work effectively on a portable MinION sequencing device. Using Deeplex Myc-TB on MinION shows great potential for saving money and quickly giving important medical information. Despite the MinION having higher raw error rates, there is still valuable data available. In general, this platform can be used more widely to treat tough bacteria that resist drugs in clinical settings. Description and analysis of microbial community of diseases Microbial community plays a crucial role in diseases. taking care of human health is important. It is essential to keep the human body in balance. Changes in internal pathologic conditions and external interference can easily disrupt the microbial community. In many situations, infections in clinics can be complicated, and it's common to have more than one infection at the same time, especially in diseases affecting the respiratory and intestinal systems. Arranging samples from patients in a certain order might be better than the current ways of identifying pathogens using molecules. Sequencing can find both known and new harmful species in just one test, which is why it's important. Metagenome sequencing is a crucial method for studying and understanding different types of genetic material. In a research project, scientists examined microbial communities. Comparing the results of nasal microbiota, the focus was on the bacterial makeup of intricate microbial groups. 16S rRNA gene sequencing is utilized with both Illumina and nanopore technologies to identify different types of bacteria. The nanopore sequencing platform can detect bacterial genera in the nasal microbiota just like the Illumina platform, but the nanopore platform has its own advantages. Struggle with identifying bacteria in the *Corynebacterium* group. Another research project also found this issue. Studied if using nanopore sequencing with metagenomics is possible and accurate for examining clinical respiratory samples. The findings indicated that nanopore sequencing works well with positive microbiologic cultures to diagnose severe pneumonia. Scientists often use metagenomics, which involves studying 16S rRNA sequencing, to examine how many bacteria are in the human body and how they are connected to each other. Most of the time, sequencing small parts of the 16S rRNA gene is not effective in identifying diseases. Scientists use long-amplicon PCR-based methods with nanopore sequencing to categorize organisms into their genus and species levels. created to improve the ability to classify organisms more accurately. Two unique genetic markers were used for this purpose. We checked the length of 16S rRNA and the 16S-ITS-23S region in the *rrn* operon. We analyzed these regions by sequencing them. We studied a clinical sample of *Staphylococcus pseudintermedius*, along with two fake communities and two sets of low-biomass samples from dog skin. We focused on the 16S-ITS region. Researchers found that the 23S region of the *rrn* operon is the best option for improving resolution at the species level. Short-read technology limits the accuracy of identifying bacterial species due to the length of the reads. Sequencing methods that use next-generation technology usually amplify 1-3 hyper-variable regions. Analyze different parts of the 16S rRNA gene to find out which bacteria species they belong to by checking databases with 16S rRNA gene sequences. Considering the benefit of the extended reading

capacity of nanopore sequencing. By using different platforms, we can get longer target sequences. This helps us identify species better and be more precise. Identifying the types of microbes in a sample helps us better understand the makeup of the microbial community in a clinical sample. By using genomics and metagenomics with nanopore sequencing technologies, researchers are able to study infectious disease outbreaks more effectively. With research on the human microbiome, there are chances to develop treatments and prevent diseases caused by harmful microorganisms. The rise in numbers can greatly increase. Also, these proven techniques can be applied to a wider range of situations. Detecting different types of microbes in a complicated environment can now be done more quickly and thoroughly.

IV. RESULTS AND DISCUSSION

. Conclusion The arrival of nanopore long-read sequencing has given us a strong tool for diagnosing, investigating, and monitoring. Looking ahead, there are exciting possibilities for how this technology can be used in the future. Nanopore sequencing helps capture detailed information on infectious diseases by producing long-read sequences. A set of ordered data helps improve how pathogens are classified. This is done by using nanopore technology. Sequencing based on whole-genome is a fast and detailed way to accurately show different drug resistance patterns. Nanopore technology is crucial for quickly showing how drug resistance is spreading in bacteria. Using this method helps us understand how bacteria are evolving and becoming resistant to drugs. Scientists are now looking into using advanced techniques to study bacterial communities more extensively. However, these methods have shown some clear biases in the types of bacteria that are identified. More research and time are needed to fully understand the abundance. We must thoroughly investigate the potential of this method in identifying and treating infectious diseases in a clinical setting. It is clear that nanopore sequencing can be used for infectious diseases as well. Specialized reserve for quickly identifying pathogens during future pandemics has seen recent advancements. Advancements in sequencing technology, like SmidgION, have made it possible to use smaller devices that can still generate long reads. Nanopore sequencing technology is easy to use in the field because it is portable, inexpensive, and effective for analyzing sequences. A flexible and easy-to-use diagnostic method that works across different cultures, offering high sensitivity and accuracy. It shows potential for the future. The use of apps in clinical infectious diseases and for keeping track of health can help gather data that can be used to detect outbreaks. Genes that resist drugs are present in every technique. Using diagnostic methods has its pros and cons, so it's not always possible to swap out old molecular methods with new ones. More and more research has shown that using a combination of NGS and traditional methods can be effective. Nanopore sequencing gives more accurate results at a lower price. It is the right choice for sequencing. Protocols need to be created based on the specific goals of the research and the medical requirements

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REFERENCES

1. Simner PJ, Miller HB, Breitwieser FP, et al. Development and optimization of metagenomic next-generation sequencing methods for cerebrospinal fluid diagnostics.
2. J Clin Microbiol, 2018; 56, e00472–18. 21. Li YM, Xiu LS, Liu JW, et al. A multiplex assay for characterization of antimicrobial resistance in *Neisseria gonorrhoeae* using multi-PCR coupled with mass spectrometry.
3. J Antimicrob Chemother, 2020; 75, 2817–25. 22. Zhang C, Xiao Y, Du J, et al. Application of multiplex PCR coupled with matrix-assisted laser desorption ionization-time of flight analysis for simultaneous detection of 21 common respiratory viruses.
4. J Clin Microbiol, 2015; 53, 2549–54. 23. Zhang C, Xiu LS, Xiao Y, et al. Simultaneous detection of key bacterial pathogens related to pneumonia and meningitis using multiplex PCR coupled with mass spectrometry. Front Cell Infect Microbiol, 2018; 8, 107. 24
5. . Hernández OH, Gutiérrez-Escolano AL, Cancio-Lonches C, et al. Multiplex PCR method for the detection of human norovirus, *Salmonella* spp. , *Shigella* spp. , and Shiga toxin producing

- Escherichia coli in blackberry, coriander, lettuce and strawberry. *Food Microbiol*, 2022; 102, 103926. 25.
6. Xiu L, Li YM, Wang F, et al. Multiplex high-resolution melting assay for simultaneous identification of molecular markers associated with extended-spectrum cephalosporins and azithromycin resistance in *Neisseria gonorrhoeae*. *J Mol Diagn*, 2020; 22, 1344–55. 26
 7. . Xiu L, Zhang C, Li YM, et al. High-resolution melting analysis for rapid detection of the internationally spreading ceftriaxone-resistant *Neisseria gonorrhoeae* FC428 clone. *J Antimicrob Chemother*, 2020; 75, 106–9. 27.
 8. Naccache SN, Federman S, Veeraghavan N, et al. A cloud-compatible bioinformatics pipeline for ultrarapid pathogen identification from next-generation sequencing of clinical samples. *Genome Res*, 2014; 24, 1180–92. 28.
 9. Van Dijk EL, Auger H, Jaszczyszyn Y, et al. Ten years of nextgeneration sequencing technology. *Trends Genet*, 2014; 30, 29. 418–26.
 10. Zhao L, Zhang H, Kohlen MV, et al. Analysis of transcriptome and epitranscriptome in plants using PacBio Iso-Seq and Nanopore-based direct RNA sequencing. *Front Genet*, 2019; 10, 253. 30.
 11. Eid J, Fehr A, Gray J, et al. Real-time DNA sequencing from single polymerase molecules. *Science*, 2009; 323, 133–8. 31.
 12. Giusti B, Sticchi E, De Cario R, et al. Genetic bases of bicuspid aortic valve: the contribution of traditional and highthroughput sequencing approaches on research and diagnosis. *Front Physiol*, 2017; 8, 612. 32.
 13. Niedringhaus TP, Milanova D, Kerby MB, et al. Landscape of next-generation sequencing technologies. *Anal Chem*, 2011; 83, 4327–41. 33. Wang Y, Yang QP, Wang ZM. The evolution of nanopore sequencing. *Front Genet*, 2015; 5, 449. 34.
 14. Reuter JA, Spacek DV, Snyder MP. High-throughput sequencing technologies. *Mol Cell*, 2015; 58, 586–97. 35.
 15. Yahara K, Suzuki M, Hirabayashi A, et al. Long-read metagenomics using PromethION uncovers oral bacteriophages and their interaction with host bacteria. *Nat Commun*, 2021; 12, 27. 36.
 16. Leggett RM, Clark MD. A world of opportunities with nanopore sequencing. *J Exp Bot*, 2017; 68, 5419–29. 37. Imai K, Tamura K, Tanigaki T, et al. Whole genome sequencing of influenza A and B viruses with the MinION sequencer in the clinical setting: a pilot study. *Front Microbiol*, 2018; 9, 2748. 38.
 17. McIntyre ABR, Rizzardi L, Yu AM, et al. Nanopore sequencing in microgravity. *npj Microgravity*, 2016; 2, 16035. 39.
 18. Castro-Wallace SL, Chiu CY, John KK, et al. Nanopore DNA sequencing and genome assembly on the international space station. *Sci Rep*, 2017; 7, 18022. 40.
 19. Pennisi E. Genome sequencing. Search for pore-fection. *Science*, 2012; 336, 534–7. 41.
 20. Jain M, Koren S, Miga KH, et al. Nanopore sequencing and assembly of a human genome with ultra-long reads. *Nat Biotechnol*, 2018; 36, 338–45. 42.
 21. Badenes ML, Martí AFI, Ríos G, et al. Application of genomic technologies to the breeding of trees. *Front Genet*, 2016; 7, 198. 43.
 22. Sereika M, Kirkegaard RH, Karst SM, et al. Oxford Nanopore R10.4 long-read sequencing enables near-perfect bacterial genomes from pure cultures and metagenomes without short-read or reference polishing. *bioRxiv*, 2021. 44.
 23. Roychowdhury S, Chinnaiyan AM. Translating cancer genomes and transcriptomes for precision oncology. *CA Cancer J Clin*, 2016; 66, 75–88. 45
 24. . Arumugam M, Raes J, Pelletier E, et al. Enterotypes of the human gut microbiome. *Nature*, 2011; 473, 174–80. 46.
 25. De Vlaminc I, Khush KK, Strehl C, et al. Temporal response of the human virome to immunosuppression and antiviral therapy. *Cell*, 2013; 155, 1178–87. 47.
 26. Teng L, Lee S, Ginn A, et al. Genomic comparison reveals natural occurrence of clinically relevant multidrug-resistant extended-spectrum- β -Lactamase-producing *Escherichia coli* strains. *Appl Environ Microbiol*, 2019; 85, e03030–18. 48.
 27. Díaz-Viraqué F, Pita S, Greif G, et al. Nanopore sequencing significantly improves genome assembly of the protozoan parasite *Trypanosoma cruzi*. *Genome Biol Evol*, 2019; 11, 1952–7. 49.
 28. Moss EL, Maghini DG, Bhatt AS. Complete, closed bacterial genomes from microbiomes using nanopore sequencing. *Nat Biotechnol*, 2020; 38, 701–7. 50.

29. Stark R, Grzelak M, Hadfield J. RNA sequencing: the teenage years. Nat Rev Genet, 2019; 20, 631–56.

